RESEARCH ARTICLE | Stem Cells: Physiology and Microenvironment

Myelo-erythroid commitment after burn injury is under β-adrenergic control via MafB regulation

Shirin Hasan,1,2 Nicholas B. Johnson,1,2 Michael J. Mosier,1,2 Ravi Shankar,1,2 Peggie Conrad,1,2 Andrea Szilagyi,2 Richard L. Gamelli,1,2 and Kuzhali Muthumalaiappan1,2

1Department of Surgery, Loyola University Chicago, Health Sciences Division, Maywood, Illinois; and 2Burn and Shock Trauma Research Institute, Loyola University Chicago, Health Sciences Division, Maywood, Illinois

Submitted 17 May 2016; accepted in final form 22 December 2016

Hasan S, Johnson NB, Mosier MJ, Shankar R, Conrad P, Szilagyi A, Gamelli RL, Muthumalaiappan K. Myelo-erythroid commitment after burn injury is under β-adrenergic control via MafB regulation. Am J Physiol Cell Physiol 312: C286–C301, 2017. First published December 28, 2016; doi:10.1152/ajpcell.00139.2016.—Severely injured burn patients receive multiple blood transfusions for anemia of critical illness despite the adverse consequences. One limiting factor to consider alternate treatment strategies is the lack of a reliable test platform to study molecular mechanisms of impaired erythropoiesis. This study illustrates how conditions resulting in a high catecholamine microenvironment such as burns can instigate myelo-erythroid reprioritization influenced by β-adrenergic stimulation leading to anemia. In a mouse model of scald burn injury, we observed, along with a threefold increase in bone marrow LSK cells (lin<sup>−</sup> Sca1<sup>−</sup> cKit<sup>−</sup>), that the myeloid shift is accompanied with a significant reduction in megakaryocyte erythrocyte progenitors (MEPs). β-Blocker administration (propranolol) for 6 days after burn, not only reduced the number of LSKs and MafB<sup>+</sup> cells in multipotent progenitors, but also influenced myelo-erythroid bifurcation by increasing the MEPs and reducing the granulocyte monocyte progenitors in the bone marrow of burn mice. Furthermore, similar results were observed in burn patients’ peripheral blood mononuclear cells, ex vivo culture system, demonstrating that commitment stage of erythropoiesis is impaired in burn patients and intervention with propranolol (nonselective β1,2-adrenergic blocker) increases MEPs. Also, MafB<sup>+</sup> cells that were significantly increased following standard burn care could be mitigated when propranolol was administered to burn patients, establishing the mechanistic regulation of erythropoiesis commitment by myeloid regulatory transcription factor MafB. Overall, results demonstrate that β-adrenergic blockers following burn injury can redirect the hematopoietic commitment toward erythroid lineage by lowering MafB expression in multipotent progenitors and be of potential therapeutic value to increase erythropoietin responsiveness in burn patients.

IN THE UNITED STATES ALONE, ~12 million units of packed red blood cells (pRBCs) are transfused each year (7). Many patients in the intensive care unit (ICU) receive blood transfusions during their ICU stay. A common laboratory finding in patients in the ICU is a significant decrease in hemoglobin and hematocrit levels, leading to anemia within 72 h of admission (47) that persists for at least 6 mo even after the ICU discharge (2). One of the major consequences of burn injury is the persistent anemia in patients with >10% total burn surface area (TBSA) (33, 51). Regardless of hemoglobin triggers (23), burn patients continue to receive multiple transfusions during their hospital stay. Over half of those transfusions are unrelated to surgical procedures (36), meaning red cell production is impaired in burn patients and cannot be explained by surgical blood loss alone. Of equal importance is that multiple transfusions lead to significant morbidity and mortality in burn patients (33). Despite the adverse consequences, treatment strategies to increase red cell production and to reduce transfusions are hampered by a lack of reliable test platforms to study the molecular mechanisms of impaired erythropoiesis and to assess the efficacy of treatments other than transfusion in critically ill burn patients. Here we report one such test platform that is both diagnostic and prognostic for anemia of critical illness in burn patients.

Bone marrow is the seat of hematopoiesis in mammalian biology wherein hematopoietic stem cells (HSCs) sequentially differentiate by a predefined program to form white blood cells, RBCs, and platelets and thus help regulate immunity, efficient oxygen delivery to tissues, and coagulation (38). Compromise in any one of the hematopoietic lineages of development can lead to severe health consequences. Bone marrow dysfunction has been observed in the erythroid fraction of thermally injured patients (46). In an electron microscope study of bone marrow fragments obtained by sternal biopsy, erythropoiesis was found to be depressed compared with granulopoiesis in burned patients (25). Because bone marrow is not a readily accessible compartment in burn patients, we have established a method to use peripheral blood mononuclear cells (PBMCs) to study the commitment stage of erythropoiesis. Previously, we utilized the hematopoietic cells residing in the PBMCs to study persistent anemia in burn patients and found that the erythropoietin (Epo)-dependent proliferation stage of erythropoiesis was intact in burn patients. More specifically, colony-forming unit-erythroid (CFU-E) production decreased within a week after burn and continued to decline for a study period of 1 mo (48). Given that rHu-Epo treatment fails to augment erythropoiesis both in burn patients (29) and animal models (37), and because Epo receptors begin to be expressed from the CFU-E stage (21, 50), it is likely that erythropoietic defects are initiated upstream of CFU-Es, which is at the commitment stage. In a mouse model of scald burn (15% TBSA), we have previously shown that bone marrow hematopoiesis...
Erythropoietic commitment is skewed away from erythroid and toward myeloid cells (37) and that monocytopenia is under the influence of norepinephrine (44). Additionally, functional β2-adrenergic receptors expressed by the hematopoietic progenitor cells at sequential ranks of lineage commitments suggest an essential role for catecholamines in dictating the bone marrow myeloid cells (32). Our recent work has shown that burn injury augments monocyte differentiation via high expression of MafB (16, 49) and that MafB is increased in common myeloid progenitors (CMPs) (18). Nonetheless, what causes the activation of myeloid-specific transcription factor MafB, which orchestrates lineage preferences and inhibits the erythropoietic potential following burn injury, is not known.

Our previous studies, and that of others, have led us to hypothesize that β-adrenergic stimulation following burn injury elicits a shift in the commitment of HSCs away from the megakaryocyte erythrocyte progenitors (MEPs) and toward nonerythroid progenitors with a possible involvement of myeloid-specific transcription factor MafB. We utilized burn patients’ PBMC-derived ex vivo culture system as well as a mouse model of burn injury to investigate the defects upstream of CFU-E production at the stage of erythroid commitment while focusing on the direct role of MafB. We further probed the effect of administering propranolol (nonselective β1- and β2-blocker) after burn injury and discovered the concomitant role of MafB-expressing multipotent progenitor (MPP) population on myelo-erythroid commitment. We established this myelo-erythroid commitment pattern in bone marrow progenitors of burn mice and also in burn patients using PBMC-derived ex vivo-generated MPPs. Results reveal that early-stage Epo-independent myelo-erythroid commitment is orchestrated via β1- and β2-adrenergic mechanisms following burn injury through MafB regulation.

**MATERIALS AND METHODS**

**Human Blood Samples**

Following institutional review board approval, we enrolled fifteen adult patients of both sexes, older than 18 yr, with >20% TBSA burn admitted between 2013 and 2016. Patients with chemical burns, electrical burns, anoxic brain injury, self-inflicted burn, pregnancy, and lactation were excluded. For those patients who received transfusions, we waited at least 12 h after the leuco-reduced pRBCs transfusions before the blood samples were drawn for the study. Out of the fifteen enrolled, thirteen patients satisfied the inclusion criteria, and informed, written consent was received. Of the two patients who were excluded from the study, one did not follow the transfusion criteria (samples were drawn before 12 h following transfusion), and the other was already on some other prescription medication before and after admission.

Blood samples were collected and analyzed at four different time points, 1–3 days, 7–10 days, 30–35 days, and 42–48 days after burn.

**Results**

Results from first and second samples were grouped together to represent data at <2 wk; results from the third and fourth samples were grouped together to represent data at 4–7 wk. Blinded analysis revealed a clear demarcation of the results between patients after follow-up. Further stratification based on treatment module examination of each patient revealed that some patients happened to receive β-adrenergic blocker propranolol during the course of burn care as part of another clinical trial. Some (8/13) patients were given propranolol to reduce heart rate as part of a different prospective ongoing study. We then divided the burn patients into two cohorts, standard burn care patients (SBC) and those who received propranolol during their burn care. Patient demographics are shown in Tables 1 and 2. Propranolol dosage regimen in burn patients is provided in Table 2. For baseline values, blood samples from eight matched healthy volunteers (Table 3) were drawn and analyzed with informed, written consent.

**Isolation of Peripheral Blood Mononuclear Cells**

Blood samples from patients and control subjects were collected in BD Vacutainer CPT cell preparation tubes (BD Biosciences, Franklin Lakes, NJ). PBMCs were isolated by Ficoll-Hypaque density-gradient centrifugation (11, 24). Samples of PBMCs (4 × 10^6 cells/ml) were stored at −80°C in fetal bovine serum with 10% dimethyl sulfoxide (Sigma-Aldrich, St. Louis, MO) until analysis.

**Ex Vivo Erythroid Progenitor Differentiation From PBMCs**

Ficoll separated PBMCs were placed in a growth factor cocktail conducive to preserve and proliferate the residing HSCs and differentiate into myeloid-erythroid progenitors as described previously for phase 1 culture (48). Briefly, PBMCs (1 × 10^6/ml) were cultured in serum-free expansion medium (Stem Cell Technologies, Vancouver, BC, Canada) supplemented with cyclosporine A (1 ng/ml; Sigma-Aldrich), granulocyte macrophage colony-stimulating factor (20 ng/ml; eBioscience, San Diego, CA), stem cell factor (45 ng/ml; Stem Cell Technologies), and interleukin-3 (5 ng/ml; Affymetrix, Santa Clara, CA). Cells were then incubated at 37°C with 5% CO2 for 5 days.

**Enrichment of Lineage-Negative Cells by Magnetic Isolation and Flow Cytometric Sorting**

On day 6, an aliquot of nonadherent cells was counted by Trypan blue exclusion, and the rest were washed in PBS containing 10% (vol/vol) FBS, and lineage-negative (lin.neg) cells were enriched using magnetic separation. Lineage-positive cells were excluded by staining with biotin-conjugated antibody cocktail specific for the lineage antigens (CD3, clone OKT3, cat. no. 13-0037-80; CD14, clone 61D3, cat. no. 13-0149-82; CD19, clone HIB19, cat. no. 13-0199-80; and CD20, clone 2H7, cat. no. 13-0209-80) (eBioscience) for 15 min at 4°C. After extensive washing with MACS buffer (PBS, 0.5% (wt/vol) BSA, 0.1% (wt/vol) n-glucose, 0.09% (wt/vol) sodium azide, and 2 mM EDTA), cells were incubated with anti-biotin magnetic microbeads (Miltenyi Biotec, Auburn, CA) for 15 min at 4°C. Cells were then washed and resuspended in 1 ml of MACS buffer. Magnetic cell separation was carried out using the AutoMACS separator (Miltenyi Biotec) by referring to the AutoMACS User Manual and applying the

**Table 1. Demographics of standard burn care patients**

<table>
<thead>
<tr>
<th>Patient</th>
<th>Sex</th>
<th>Age, yr</th>
<th>TBSA, %</th>
<th>Transfusions, pRBC Units</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patient 1</td>
<td>M</td>
<td>44</td>
<td>67</td>
<td>33.5</td>
</tr>
<tr>
<td>Patient 2</td>
<td>M</td>
<td>43</td>
<td>23</td>
<td>None</td>
</tr>
<tr>
<td>Patient 4</td>
<td>M</td>
<td>26</td>
<td>29</td>
<td>None</td>
</tr>
<tr>
<td>Patient 8 (deceased)</td>
<td>F</td>
<td>59</td>
<td>50</td>
<td>24</td>
</tr>
<tr>
<td>Patient 15</td>
<td>F</td>
<td>26</td>
<td>35</td>
<td>18</td>
</tr>
</tbody>
</table>

M = 3; F = 2; Mean ± SE: 39.6 ± 6.2; Mean ± SE: 40.8 ± 7.9; Mean ± SE: 15.1 ± 6.6; Median = 18

TBSA, total burn surface area; pRBCs, packed red blood cells; M, male; F, female.
separation program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.

The enriched lin\(^{-}\) fraction (the eluate) was stained with cell surface antigens using anti-CD34-APC (clone-561, cat. no. 343608; BD PharMingen, San Jose, CA), anti-CD123-PE.Cy7 (clone 555462; BD PharMingen, San Jose, CA), anti-CD45RA-V450 (clone 555462; BD PharMingen, San Jose, CA), and anti-CD34-APC (clone 555462; BD PharMingen, San Jose, CA) antibodies. An aliquot was separated program “depletes,” which retains lin\(^{-}\) cells in the magnetic column.
with the following biotin-conjugated lineage-specific primary antibodies: anti-CD86 (clone GL1, cat. no. 553690), anti-CD11c (clone HL3, cat. no. 553800), anti-Ter119 (clone Ter119, cat. no. 553672), anti CD19 (clone1D3, cat. no. 553784), anti-CD11b (clone M1/70, cat. no. 553029), anti-Gr1 (clone RB6–8C5, cat. no. 553125), anti-CD127 (clone A7R34, cat. no. 13-1271-82), and anti-CD3e (clone 145-2C11, cat. no. 553060)
(BD Biosciences) followed by incubation with anti-biotin magnetic beads (Miltenyi Biotec). Magnetic cell separation was carried out using the AutoMACS separator (Miltenyi Biotec) referring to the AutoMACS User Manual and applying the separation program “depletes.” The enriched lineage-negative (linneg) fraction was surface stained with PerCP-Cy5.5-CD11b (BD Biosciences; clone M1/70, cat. no. 552850) antibodies for Ter119 (eBioscience; clone Ter119, cat. no. 45-5921-82) and Pe.Cy7-linnegSca1 (LSK) cells were sorted from TBM cells as described above. Total RNA was isolated from LSKs using RNeasy Mini Kit (Qiagen, Valencia, CA) as described by the manufacturer. First-strand cDNA was synthesized from 180 ng total RNA using a High-Capacity cDNA Reverse Transcription Kit (Life Technologies, Carlsbad, CA), and reactions were run on a Veriti 96-well Fast Thermalcycler (Life Technologies) per the manufacturer’s instructions. The expression levels of colony-stimulating factor 1 receptor (CSF1R) and colony-stimulating factor 2 receptor β common subunit (CSF2RB) were analyzed by real-time PCR using TaqMan primer and probe sets and TaqMan Fast Advanced Master Mix (Life Technologies) in StepOnePlus Real-Time PCR System (Applied Biosystems, Carlsbad, CA). GAPDH was included as a normalization control for each reaction. Target gene Ct cycle values were normalized to GAPDH Ct value, and fold gene expression in burn group is expressed relative to the sham group.

**Immunohistochemical Analysis of Tyrosine Hydroxylase**

Brains from sham and burn mice (PBD7) were freshly isolated and snap frozen in liquid nitrogen and stored at −80°C until analysis. Tissue lysates were prepared by homogenization in RIPA buffer containing Halt protease inhibitor cocktail (Pierce, Rockford, IL) and sonication, and lysates were then obtained after centrifugation. Protein concentrations were determined using Pierce BCA-200 Protein Assay kit, 5 (propranolol) as explained above for MafB-expressing cells. Representative FACS plots of patient 2 (SBC) and patient 5 (propranolol) as explained above for MafB-expressing cells.

**Confocal Microscopy**

An aliquot of MafB-stained MPPs was cytospun onto microscopic slides and preserved using Vectashield H-1500 mounting medium with DAPI (Vector Laboratories, Burlingame, CA). A Zeiss LSM 510 laser-scanning microscope (Carl Zeiss MicroImaging, Jena, Germany) was used to view with C-Apochromat 40x/1.20 water immersion, and ×40 images were acquired using Zeiss LSM 510, version 4.2, SP1 software.

**Bone Marrow Mature Myeloid and Erythroid Cells**

An aliquot (1 × 10⁶) of TBM cells was labeled with PerCP.Cy5.5-Ter119 (eBioscience; clone Ter119, cat. no. 45-5921-82) and PeCy7-CD11b (BD Biosciences; clone M1/70, cat. no. 552850) antibodies for 30 min at 4°C in the dark. Cells were washed and resuspended in PBS and immediately analyzed with a FACS Canto II (BD Biosciences).

**Total RNA Isolation, cDNA Synthesis, and Quantitative PCR**

Lin⁰Scal¹cKit⁺ (LSK) cells were sorted from TBM as described above. Total RNA was isolated from LSKs using RNeasy Mini Kit (Qiagen, Valencia, CA) as described by the manufacturer. First-strand cDNA was synthesizes from 180 ng total RNA using a High-Capacity cDNA Reverse Transcription Kit (Life Technologies, Carlsbad, CA), and reactions were run on a Veriti 96-well Fast Thermocycler (Life Technologies) per the manufacturer’s instructions. The expression levels of colony-stimulating factor 1 receptor (CSF1R) and colony-stimulating factor 2 receptor β common subunit (CSF2RB) were analyzed by real-time PCR using TaqMan primer probes and TaqMan Fast Advanced Master Mix (Life Technologies) in StepOnePlus Real-Time PCR System (Applied Biosystems, Carlsbad, CA). GAPDH was included as a normalization control for each analysis. Target gene Ct cycle values were normalized to GAPDH Ct values. Data were calculated using the ΔΔCt method, and fold gene expression in burn group is expressed relative to the sham group.

**Statistical Analysis**

Results from all experiments are expressed as means ± SE. All experiments were repeated at least three times. The number of animals used per experiment is given in respective figure legends. Analysis of variance with Tukey’s post hoc test or Student-Newman-Keuls multiple-comparison test using KaleidaGraph statistical program (version 4.1.0, Synergy Software, Reading, PA) were carried out for multiple comparisons. Statistical significance was set at P < 0.05.

**RESULTS**

**PBMC-Derived Hematopoietic Progenitors**

PBMC resident HSCs were differentiated into myelo-erythroid progenitors as described previously (48). Nonadherent cells collected from culture plates were subjected to flow cytometric categorization based on lineage⁰CD34⁺CD38⁻, which are MPPs. MPPs were subdivided according to differential expressions of intracellular MafB. MPPs were next further classified into nonerythroids and MEPs based on a combination of cell surface antigens. MEPs express neither CD123 nor CD45RA (CD123⁰CD45RA⁰), whereas nonerythroids are gated as CD123⁺/− CD45RA⁺ as described previously (30). Detailed gating strategies are shown in Fig. 1A. Representative FACS plots of patient 2 (SBC) and patient 5 (propranolol) as explained above for MafB-expressing cells.
in MPPs and percentages of MEPs and nonerythroids are shown in Fig. 1B.

**Propranolol Treatment in Burn Patients Can Restore PBMC-Derived MEPs**

We have previously shown that Epo-dependent erythropoiesis is not affected in burn patients and that CFU-E is probably the rate-limiting entity, affecting erythropoiesis (48). In this study, PBMC-derived MEPs were determined and expressed as percentages of MPPs. PBMC-derived MEPs of SBC patients and propranolol-treated patients are shown in Fig. 2, A and B, respectively. The percentage of MEPs was significantly decreased at <2 wk post burn in SBC patients compared with controls and continued to remain significantly low till 4–7 wk (P < 0.01 vs. controls). In patients who received propranolol, the decrease in PBMC-derived MEPs was not significantly different from controls but was significantly higher than SBC patients at <2 wk (P < 0.05 vs. SBC) and 4–7 wk post burn (P < 0.001 vs. SBC; Fig. 2C). Total MPP cell counts from cell culture plates are shown as bar graphs in Fig. 2D, which indicated robust proliferation of hematopoietic stem and progenitor cells.

**Propranolol Treatment in Burn Patients Decreases MafB Expression in PBMC-Derived MPPs**

Our recent study has shown that burn injury-mediated increase in myeloid transcription factor MafB restricts the erythroid potential of bone marrow CMPs (18). To validate whether β-adrenergic signaling is the primary regulator of MafB controlling the myelo-erythroid commitment in burn patients, we quantified MafB-expressing cells in the ex vivo PBMC-derived MPPs. Percentages of MafB^+ MPPs in SBC patients and propranolol-treated patients are shown in Fig. 3, A and B, respectively. Compared with controls, the percentages of MafB^+ cells in the MPPs were significantly increased in patients with SBC at <2 wk (P < 0.01) and

---

**Fig. 2. Ex vivo PBMC-derived MEPs.** PBMC-derived lin^- CD34^- CD38^- cells are signified as MPPs. Line graph y-axes represent percentage of MEPs in PBMC-derived MPPs, and x-axis represents weeks post burn from week 1 to week 7. Each line in the graph represents data for each patient included in the study, and the dashed line represents baseline values (means ± SE) from control PBMCs. A: black lines represent percentage of MEPs in SBC-treated patients. B: gray lines represent PR-treated patients. The numbers next to the lines represent each patient matching those in Table 1. C: bar graph representing means ± SE of MEPs calculated as percentage of MPPs. D: bar graph representing proliferation of PBMC-derived MPPs in ex vivo culture condition in the presence of SCF, Hu-GMCSF, Hu-IL-3, and cyclosporin A. The y-axis represents total number of MPPs per million of PBMCs seeded. Control (n = 8), SBC (n = 5), PR (n = 8). * vs. controls, ** vs. SBC. ***P < 0.001, **P < 0.01, *P < 0.05 by one-way ANOVA.
continued to remain significantly elevated till 4–7 wk post burn \((P < 0.05)\). However, in propranolol-treated patients, the percentage of MafB expressing cells decreased considerably by 4–7 wk post burn compared with SBC patients \((P < 0.05; \text{Fig. 3C})\). The mean fluorescent intensity (MFI) of MafB expression in MPPs was also significantly increased in SBC patients compared with controls \((P < 0.001)\) and was significantly reduced in propranolol-treated patients \((P < 0.001 \text{ vs. SBC})\) at all time points studied (Fig. 3D). Overall, results indicate that MafB expression correlates reciprocally with the percentage of MEPs in PBMC-derived MPPs and is regulated by \(\beta\)-adrenergic mechanisms. More importantly, MafB appears to be a negative regulator averting MEP production in burn patients and thereby orchestrating the commitment stage of hematopoiesis through \(\beta\)-adrenergic mechanisms.

Interestingly, patient 6 and patient 14 were on a regimen of propranolol, which exceeded 40–70 mg in a single dose, compared with others who received not more than 10–30 mg at one dose. These two patients did not respond as well, exhibiting lower MEPs (Fig. 2B) and higher MafB (Fig. 3B) levels. Detailed dose-response studies are warranted to draw more meaningful inferences. Nonetheless, all patients were included in the bar graphs shown as means \(\pm SE\) in this study (Figs. 2C and 3C).

**TBSA, Age, and Hemoglobin**

Percentage of TBSA did not influence percentage of MEPs either in SBC or propranolol-treated patients at <2 wk and at 4–7 wk post burn (Fig. 4A). Similarly, age of the patients had no correlation with percentage of MEPs at both time points. Data shown are at 4–7 wk post burn (Fig. 4B). Blood hemoglobin levels were retrospectively recorded

\[\text{AJP-Cell Physiol} \quad \text{doi:10.1152/ajpcell.00139.2016} \quad \text{www.ajpcell.org}\]
from the charts at the single time of sample collection for most burn patients, and the mean value (in the case of multiple recordings on a single day) was calculated and given at 2 wk and 4–7 wk post burn. Mean hemoglobin levels were below reference range (4) for the entire study period in both groups (Fig. 4C).

**Sympathetic Activity Is Increased After Burn Injury**

One way to evaluate the extent of sympathetic activity is to measure TH, which is the rate-limiting enzyme in catecholamine biosynthesis (8). Compared with sham, a 15% TBSA burn injury increased TH levels as detected by immunohistochemistry staining of brain sections at PBD7 (Fig. 5A) and the protein expression of TH by Western blot in brain tissue lysates (Fig. 5B). GAPDH served as a loading control. Bar graphs in Fig. 5C represent significantly higher TH expression relative to GAPDH following burn injury ($P < 0.01$ vs. sham).

**Burn Injury Augments CSF1R and CSF2RB Gene Expression in LSKs**

To get some insight whether the burn-induced microenvironment can influence the hematopoietic myeloid commitment, we performed gene expression analysis of myeloid-related genes, specifically *CSF1R* and *CSF2RB* in LSKs from sham and burn mice. Real-time PCR data demonstrate a significant increase ($P < 0.001$) in gene expression of *CSF1R* (4.9-fold) and *CSF2RB* (3.9-fold) in LSKs from burn mice on PBD7 compared with sham (Fig. 6, A and B, respectively). Although this observation supports the myeloid bias and a robust hematopoietic proliferation capacity after burn injury, with the assumption that the PBMC-resident HSCs are orchestrating lineage preference in burn patients, the true picture of the hematopoietic paradigm shift will emerge only from probing the bone marrow. Although bone marrow is the primary seat of erythropoiesis, obtaining bone marrow aspiration from burn patients is a highly invasive procedure. Therefore, we pursued a practical approach with a mouse model of burn injury such that the lineage-committed progenitors can be directly isolated from bone marrow with specific cell surface markers using the latest techniques.

**Propranolol Reverses Burn Injury-Mediated Increase in LSKs**

We administered propranolol subcutaneously for 6 days after burn injury to evaluate the β-adrenergic influence on burn injury-mediated increase in the LSK cell population. As seen in
the bar graph in Fig. 7A, the percentage of LSKs within the bone marrow begins to increase from as early as PBD3, and blocking with propranolol for 6 days post burn significantly lowered the proportion of LSKs expressed in lin<sup>neg</sup> bone marrow fraction (P < 0.01 vs. burn).

**Propranolol Treatment Reduces MafB-Expressing Cells in the Bone Marrow Hematopoietic MPPs (Murine: lin<sup>neg</sup> cKit<sup>+</sup>)**

We next isolated MPPs residing within the bone marrow of mice and examined the intracellular MafB expression. Overall, the MFI of MafB expression was increased in MPPs from burn mice compared with sham and was mitigated with propranolol administration as determined by flow cytometry (Fig. 7B; P < 0.05).

With confocal imaging, we observed that, among bone marrow MPPs, MafB-expressing cells were particularly increased in burn compared with sham mice, which was significantly reduced with propranolol treatment, similar to propranolol-treated burn patient PBMC-derived MPPs. Representative confocal images of MPPs that display the differential expression of MafB<sup>+</sup> cells between the three groups are shown in Fig. 7C.

We then evaluated the proportion of MafB<sup>+</sup> cells in the MPPs from total bone marrow over the course of burn. The bar graphs in Fig. 8A represent the percentage of MafB<sup>+</sup> cells (means ± SE) in all three groups harvested on PBD3, PBD7, and PBD14. We noticed a significant increase in the percentage of MafB<sup>+</sup> cells in burn mice from PBD3 (P < 0.01 vs. sham). Propranolol treatment, on the other hand, significantly reduced the percentage of MafB<sup>+</sup> cells at PBD7 (P < 0.01 vs. burn saline). No significant difference was seen between propranolol and saline administration in sham mice (results not shown). To confirm whether the bone marrow effect is a continuous response to burn injury, and to control the dose of propranolol to match clinical situations, we next placed alzet pumps under the skin to deliver either propranolol or placebo continuously for 13 days.
until harvest on PBD14 (3, 15). Results from all three groups with alzet implants also indicate that the bone marrow myeloid transcription factor MafB is under β-adrenergic regulation following burn injury ($P < 0.05$ vs. burn saline pump).

Propranolol Treatment Influences Myelo-Erythroid Bifurcation in the Bone Marrow

We noticed a significant reduction in MEPs within the MPP population in the burn mice starting from PBD3, which remained low compared with sham until PBD14 (Fig. 8B). Propranolol-treated mice exhibited a significant increase in the MEP population from PBD7, which persisted until PBD14 ($P < 0.05$). In corroboration with our earlier results from burn patient PBMC-derived ex vivo culture, a similar trend in the commitment bias of MPPs away from MEPs was found in the burn mice that is counteracted with propranolol administration.

Unlike in the contrived ex vivo culture system explained previously, our in vivo mouse model will allow us to investi-
gate the counterregulatory myeloid arm of hematopoietic progenitors, particularly the GMP population. As expected, bone marrow GMP fraction was significantly increased after burn injury from PBD3 and persisted until PBD14 ($P < 0.01$ vs. sham). Daily injections of propranolol for 6 days significantly reduced GMPs to sham levels ($P < 0.01$ vs. burn); however, a subcutaneous implant with lower-dose propranolol had an insignificant effect (Fig. 8C).

**Bone Marrow Epo Responsiveness Is Improved After β-Adrenergic Blockade Following Burn Injury**

To investigate whether or not the effect of propranolol is reflected on mature lineage-committed myeloid and erythroid cells, we next evaluated the percentage of terminal lineage markers CD11b and Ter119 in the total bone marrow cells. Although there was no change between the three groups on PBD3, we noticed a significant increase in CD11b$^+$ cells and a corresponding decrease in Ter119$^+$ cells in the burn group on PBD7 (Fig. 9A). However, in the propranolol group, we noticed a reciprocal reversal in the respective myeloid and erythroid cells compared with burn mice, indicating significant progress in erythroid maturation with propranolol treatment ($P < 0.05$). Given the influence of propranolol on erythro-myeloid commitment, these results proportionately reflect those of bone marrow progenitors (GMPs and MEPs) following burn injury. Nonetheless, Epo responsiveness was also improved after propranolol administration. Visual representation of pelleted total bone marrow cells (Fig. 9B) clearly depicts the red cell mass that is almost depleted in burn, improved with propranolol, and considerably enhanced with Epo administration. Quantitative analysis by flow cytometry using cell surface markers Ter119 and side scatter is represented in sample dot plots, and the results are enumerated (means ± SE) in Table 4. However, burn injury resulted in a concurrent increase in CD11b$^+$ total bone marrow cells, which was reduced by propranolol and remained unchanged with Epo, indicating the specific response of Epo on Ter119-expressing erythroid cells. Therefore, a combined therapy with Epo to complement the action of propranolol can be effective in restoring bone marrow erythropoiesis in burn patients.

**DISCUSSION**

In this study, we examined human PBMC-derived ex vivo cultures as a proxy to bone marrow hematopoiesis by evaluating the distribution of erythroid and nonerythroid progenitors and measuring intracellular MafB expression in MPPs. We report that impaired erythropoiesis in burn patients is primarily attributable to defective Epo-MafB expression in MPPs. The possible molecular mechanism orchestrating the myeloid bias is the transcription factor MafB mediated by β1/β2-
adrenergic stimulation following burn injury. Furthermore, we confirmed this observation using an in vivo mouse model of scald burn by investigating bone marrow stem and progenitor cell distribution with and without β1/β2-adrenergic blockade and showed that bone marrow Epo responsiveness can be improved. We also established that burn-induced sympathetic stimulation is evident in a 15% TBSA mouse model.

Given the multifactorial etiology of anemia in critically ill burn patients, the only treatment option is blood transfusion, which is often associated with adverse effects (23, 33). To

### Fig. 9.

A: propranolol administration for 6 days after burn influences mature myeloid and erythroid cells in the bone marrow; y-axis: number of CD11b⁺ (×10⁶) and Ter119⁺ (×10⁶) cells per million TBM cells collected from bilateral femurs; x-axis: time course after burn, PBD3 and PBD7. Propranolol was dosed at 1.2 mg/mouse per day, delivered via subcutaneous injections. Bar graphs represent means ± SE, containing 6 mice/group and repeated 4 times. *P < 0.05, ***P < 0.001; a vs. sham; b vs. burn by one-way ANOVA. B: erythropoietin (Epo) responsiveness is improved with exogenous Epo in propranolol-treated burn mice. Pelleted total bone marrow cells from sham, PBD7, PBD7 + PR, and PBD7 + PR + Epo groups are shown along with representative dot plots showing Ter119⁺ cells in the far right quadrant (red) and CD11b⁺ cells in the upper left quadrant (blue).

### Table 4. Bone marrow myelo-erythroid cell distribution and erythropoietin responsiveness to propranolol administration after burn injury

<table>
<thead>
<tr>
<th>Total Bone Marrow Cell Distribution</th>
<th>Sham</th>
<th>PBD7</th>
<th>PBD7 + PR</th>
<th>PBD7 + PR + Epo</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ter119⁺ (erythroid)</td>
<td>33.0 ± 2.0%</td>
<td>9.0 ± 1.0%*</td>
<td>18.0 ± 0.8%*†</td>
<td>24.0 ± 0.8%*‡</td>
</tr>
<tr>
<td>CD11b⁺ (myeloid)</td>
<td>36.0 ± 2.0%</td>
<td>66.0 ± 2.0%*</td>
<td>55.0 ± 1.5%*†</td>
<td>55.0 ± 1.2%*</td>
</tr>
</tbody>
</table>

Values are means ± SE. Experiments repeated 3 times with n = 6 mice/group. Improved erythropoietin (Epo) responsiveness was observed with exogenous Epo in propranolol-treated burn mice. *P < 0.0001 vs. Sham; †P < 0.0001 vs. PBD7; ‡P < 0.008 vs. PBD7 + PR by one-way ANOVA.
consider alternate treatment strategies, we need a clearer understanding of the molecular mechanisms of anemia of critical illness in burn patients. Recent studies imply impaired bone marrow erythropoiesis as a plausible reason for anemia following burn injury (37) with a limitation in CFU-E production (48) explaining Epo resistance in burn patients (29) and the mouse model (37). To better understand the mechanism behind Epo-resistant anemia, it is logical to probe the stage upstream of CFU-Es. As burst-forming unit-erythroblasts (BFU-Es) cannot be morphologically identified, we chose to study the distribution of MEPs, the earliest identifiable erythroid progenitors at the bifurcation of CMPs to erythroid and myeloid lineages (1).

The reduction in PBMC-derived MEPs (31, 48) was consistent in all SBC patients, which can be explained by a reprioritization in the commitment of the progenitor cells or the HSCs, which is in line with our earlier studies on the mouse model of burn injury (18, 37). One of the limitations of our ex vivo culture system is the likelihood of apoptosis and compromised proliferation of MPPs in burn patients. However, a concomitant increase in nonerythroid cells with no significant differences in MPPs supports reprioritization of lineage commitment over apoptosis. Moreover, these results were consistent with bone marrow MEP and GMP distribution in our mouse model.

It is known that myelopoietic commitment of bone marrow progenitors is modulated by norepinephrine following thermal injury with sepsis (44), and now we provide additional evidence that sympathetic activation is evident in our mouse model of 15% TBSA scald burn (Fig. 5). Therefore, bone marrow myeloid shift represented by increased GMPs and CD11b+ cells can be attributed to catecholamine action following burn injury. Because elevated catecholamine levels are evident in patients over 30% TBSA burn (22) and HSCs and progenitors express β-adrenergic receptors (32), it is reasonable to believe why burn patients who were treated with propranolol during burn care (10–30 mg) showed a reciprocal reversal in nonerythroid lineage commitment. The nonerythroid fraction should predominantly represent myeloid cells based on specified culture conditions.

For an uncommitted stem cell, the expression of receptors for the essential cytokines (growth factors) is an important requisite for commitment. Moreover, the predominance of one cytokine/receptor combination over the other can influence the lineage fate of the HSCs (52). CSF2RB is a colony-stimulating factor receptor sharing a common β-subunit for granulocyte macrophage colony-stimulating factor, IL-3, and IL-5. Together, all three growth factors upon binding to CSF2R can activate cell survival, proliferation and differentiation pathways (10, 35). Of significance, monocyte progenitors from thermally injured and septic mice revealed an increase in receptors for macrophage colony-stimulating factor (M-CSF) with a consequent shift to monocytopoiesis (39). CSF1R is the receptor for M-CSF, which is also closely associated with MafB (18). We have previously documented that increase in transcription factor MafB after burn injury was, not only responsible for monocytopoiesis (16, 49), but also accountable for hematopoietic reprioritization at the commitment stage of CMPs (18). Furthermore, retroviral expression of MafB cDNA in human CD34+ hematopoietic progenitors was found to induce monocyte commitment (12). Similarly, overexpression of MafB was found to inhibit erythropoiesis through downregulation of transferrin receptor gene in yeast and the erythroblast cell line (41). In the present study, burn patients who received propranolol showed a concomitant reduction both in proportion and intensity of MafB expression in PBMC-derived MPPs. These results strongly support the reciprocal relation between MafB expression and erythroid progenitors in burn patients and further confirm that erythro-myeloid commitment is orchestrated by β1/β2-mediated response following burn injury.

The quiescent HSCs along with their capacity to maintain “stemness,” also have the ability to respond to injury and initiate a program of rapid proliferation. Moreover, bone marrow is innervated by sympathoadrenergic efferent nerve fibers (6). Sympathoadrenergic signals and chronic stress exposure are known to regulate hematopoietic stem and progenitor cell mobilization from bone marrow (14, 19). Bible et al. (5) have shown that propranolol administration in trauma patients reduced hematopoietic progenitor cell mobilization in the peripheral blood. Catecholamines particularly elevated noradrenaline levels after stress has been associated with proliferation of HSCs in the bone marrow (14). In the present study, the increase in LSKs after burn injury can be attributed to the increased sympathetic activity as evidenced by increase in TH levels in the brain, influencing the HSC niche in the bone marrow (4, 6). A similar result on β-adrenergic blockade was found by Spiegel et al. (42), when epinephrine administration to mice increased LSK cells in bone marrow, whereas propranolol treatment reduced this population. Moreover, in a dose-response relationship study, increasing doses of exogenous norepinephrine administered to uninjured rats via alzet pumps was found to decrease the generation of CFU-E and BFU-E colonies from TBM cells (34), supporting our burn model that reinforces adrenergic influence on MEP generation in mice as well as in burn patients. Moreover, the requirement of a critical threshold of adrenergic activation for normal erythropoiesis observed in the same study may explain the differential responses in MEPs to low-dose (10–30 mg) and high-dose (40–70 mg) propranolol treatments in burn patients. Nonetheless, why chemical sympathectomy with 6-OHDA seemed to have the same effect as norepinephrine administration in rats was not clear from the study of Penn et al. (34). Unlike propranolol that blocks the action of catecholamines, 6-OHDA is known to ablate tissue norepinephrine content by preventing the release of norepinephrine from nerve endings (axotomy) (44). On the basis of our present observations, we can speculate that, if adrenergic stimulation increases bone marrow LSKs, then 6-OHDA will hamper the production of LSKs, which in turn can affect the downstream BFU-Es and CFU-Es.

Propranolol-mediated increase in MEPs after burn is positively reflected in erythroblast development as Ter119 begins to be expressed on the cell surface from proerythroblasts onward to mature erythrocytes (20). Epo binding to its receptors is crucial for definitive erythropoiesis (26); we and others (9, 28, 37, 43) have shown that burn injury results in hypersensitiveness to Epo attributable to a reduction in Epo-responsive cells affecting RBC production. Our previous study using similar PBMC-derived culture technique indicated a restriction in CFU-E generation and not in Epo-dependent proliferation in burn patients (48). In line with these observations, we now show Epo-independent commitment to MEPs, which is upstream of the CFU-E stage of erythropoiesis and is impaired in burn patients. The ultimate read out to validate the efficacy of
propranolol in improving erythropoiesis is peripheral blood hemoglobin levels and the associated transfusion requirements, which are based on hemoglobin trigger. In the present study, hemoglobin levels did not change between the two groups at the time points studied, which may not truly reflect improved erythropoiesis because burn patients were followed only for 7 wk, which is one of the limitations. Similarly, although transfusion requirements trend toward less pRBC units with propranolol treatment, further stratification based on the number of operative procedures and length of stay is warranted for conclusive interpretation of implications in patient care. Another limitation is that the megakaryocyte vs. erythrocyte potential of propranolol-generated MEPs has not been tested in the present study. However, this can be ruled out by the specific action of exogenous Epo resulting in increased Ter119 cells; this further strengthens the action of propranolol in improving erythrolineage commitment of bipotential MEPs. The fact that exogenous Epo did not affect the CD11b fraction in bone marrow validates that myel-o-erythroid bifurcation is orchestrated by β1/β2-adrenergic mechanisms through MafB. Other models of psychological stress with sympathetic activity resulting in elevated catecholamine also demonstrate similar effect with propranolol in lowering CD11b⁺ cells (13, 17). Together, these results imply the role of catecholamines in myelo-erythroid commitment consequent to stressful and traumatic pathological conditions.

β-Adrenergic stimulation following burn damps bone marrow erythropoiesis, reducing MEP production by regulating MafB expression. β-Blockade in burn patients can be efficacious as a potential therapeutic tool to mitigate MafB and restore the myelo-erythroid balance. Therefore, sequential intervention with Epo could be used to supplement the effect of propranolol and boost erythroblast proliferation in efforts to reduce transfusion requirements and rescue burn-induced anemia in severely burned patients otherwise hyporesponsive to recombinant human Epo (9, 29). A schematic diagram representing the mechanistic alterations in myelo-erythroid commitment pattern leading to RBC deficits after burn injury and rescue efforts with β-blockade is shown in Fig. 10.

Overall, our work provides important mechanistic clues and an alternative platform to study bone marrow erythropoiesis in burn patients. Obtaining bone marrow from burn patients is a barrier, but our innovative application of the culture system from PBMCs allows us, not only to evaluate the mechanisms that dictate erythroblast production in burn patients, but also to pave the way for translational research and the development of a test platform for future hematopoiesis-related drug efficacy studies. However, we acknowledge that it is a small sample size, and a larger patient population-based study is essential to further strengthen our results.

GRANTS

This work was supported by National Institutes of Health (NIH) Grant R01DK097760-01 (to K. Muthumalaiappan) and NIH Training Grant T32 GM008750 (to R. Gamelli).

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.
AUTHOR CONTRIBUTIONS

S.H., N.B.J., P.C., and A.S. performed experiments; S.H., N.B.J., and K.M. analyzed data; S.H., N.B.J., and K.M. interpreted results of experiments; S.H. and K.M. prepared figures; S.H. drafted manuscript; M.J.M., R.S., R.L.G., and K.M. edited and revised manuscript; M.J.M., R.S., R.L.G., and K.M. approved final version of manuscript; K.M. conceived and designed the research.

REFERENCES


