Epoxyeicosatrienoic acids are part of the VEGF-activated signaling cascade leading to angiogenesis


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Cytochrome P-450 (CYP) epoxygenases metabolize arachidonic acid to epoxyeicosatrienoic acid (EET) regioisomers, which activate several signaling pathways to promote endothelial cell proliferation, migration, and angiogenesis. Since vascular endothelial growth factor (VEGF) plays a key role in angiogenesis, we assessed a possible role of EETs in the VEGF-activated signal transduction cascade. Stimulation with VEGF increased CYP2C promoter activity in endothelial cells and enhanced CYP2C8 mRNA and protein expression resulting in increased intracellular EET levels. VEGF-induced endothelial cell tube formation was inhibited by the EET antagonist 14,15-epoxyeicosa-5(Z)-enoic acid (14,15-EEZE), which did not affect the VEGF-induced phosphorylation of its receptor or basic fibroblast growth factor (bFGF)-stimulated tube formation. Moreover, VEGF-stimulated endothelial cell spreading in a modified spheroid assay was reduced by CYP2C antisense oligonucleotides. Mechanistically, VEGF stimulated the phosphorylation of the AMP-activated protein kinase (AMPK), which has also been linked to CYP induction, and the overexpression of a constitutively active AMPK mutant increased CYP2C expression. On the other hand, a dominant-negative AMPK mutant prevented the VEGF-induced increase in CYP2C RNA and protein expression in human endothelial cells. In vivo (Matrigel plug assay) in mice, endothelial cells were recruited into VEGF-impregnated plugs containing bFGF. Taken together, our data indicate that CYP2C-derived EETs participate as second messengers in the angiogenic response initiated by VEGF and that preventing the increase in CYP expression curtails the angiogenic response to VEGF.

EPOXYEICOSATRIENOIC ACIDS (EETs) are metabolites of arachidonic acid generated by cytochrome P-450 (CYP) epoxygenases. The majority of CYP enzymes are primarily expressed in the liver, but several CYP enzymes can be detected in the cardiovascular system. Of those reported to date, most is known about the cardiovascular actions of proteins belonging to the CYP4A, 2C, and 2J families. CYP4A enzymes generate the potent vasoconstrictor 20-hydroxyeicosatetraenoic acid (20-HETE), which is implicated in the regulation of myogenic tone. The 2C and 2J epoxygenases, on the other hand, generate EETs, although some enzymes can also generate 20-HETE (for review see Ref. 39).

The realization that EETs, in particular 11,12- and 14,15-EET, can activate large conductance Ca2+-activated K+ channels on vascular smooth muscle cells to elicit hyperpolarization and relaxation led to their identification as a class of “endothelium-derived hyperpolarizing factor or EDHF” (2, 13). However, although CYP epoxigenase activation and EET production is generally associated with vasodilatation, the different EETs exert a variety of membrane potential-independent effects and are now recognized as intracellular signaling molecules and have been attributed with anti-inflammatory, fibrinolytic and anti-apoptotic properties (for review see Ref 31). One of the more recently explored functions of these enzymes is their ability to promote endothelial cell proliferation (4, 16, 28, 43) as well as endothelial cell migration and degradation of the extracellular matrix (29). The mechanisms underlying these actions of the EETs have not been completely resolved but are reported to involve activation of several signaling pathways, including activation of tyrosine kinases and phosphatases (16, 22), activation of mitogen-activated protein (MAP) kinase phosphatases, and inhibition of the c-Jun NH2-terminal kinase (35) as well as transactivation of the epidermal growth factor receptor (6, 30).

The endogenous expression of CYP epoxygenases can be modulated by different stimuli, in particular, hemodynamic and physiochemical forces are thought essential to maintain CYP2C expression in endothelial cells in situ (14). Hypoxia is also an effective inducer of CYP2C8/9 expression, a mechanism that contributes to hypoxia-induced angiogenesis in human endothelial cells (29). However, nothing is known about the involvement of CYP2C epoxygenases in the signaling pathways activated by other growth factors.

The vascular endothelial growth factor (VEGF) is a key regulator of physiological and pathological angiogenesis. In vitro, VEGF induces endothelial cell proliferation and migration and is a survival factor for endothelial cells (for review see Ref. 3). Since the expression of VEGF is dependent on hypoxia (27), we postulated a link between VEGF- and CYP-epoxygenase signaling. Therefore, in the present study we assessed the effects of VEGF on CYP2C epoxygenase expression and activity in human endothelial cells. In addition, we determined the effects of interfering with CYP activity or EET levels on VEGF-induced angiogenesis in vivo.

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METHODS

Materials

11,12-EET was purchased from Cayman Chemicals (Massy, France), and growth factor-reduced basement membrane matrix (Matrigel) was from BD Biosciences (San Jose, CA). The CYP2C8 antibody was from Acris (Hidenhausen, Germany) and CYP2C8-containing bacterial supemesomes (Gentest, Woburn, MA) were used as a positive control for immunoblotting. 14,15-Epoxyeicosa-5Z,E11-ene (14,15-EEZE) and N-methylsulfonyl-6-(2-propargyloxyphenyl)-hexanamide (MS-PPOH) were synthesized as described (20). Sulfaphenazole, the antibody recognizing β-actin, and all other chemicals were from Sigma (Taufkirchen, Germany).

Cell Culture

Human umbilical vein endothelial cells (30) and murine lung endothelial cells (15) were isolated and cultured as described. The porcine aortic endothelial cell lines overexpressing VEGFR1 and VEGFR2 were kindly provided by Dr. J. Waltenberger (Maastricht, The Netherlands). The investigation conforms with the principles outlined in the Declaration of Helsinki (Cardiovasc Res 35: 2-41997). In some experiments subconfluent human endothelial cells were infected with adenoviruses to overexpress constitutively active AMP-activated protein kinase (AMPK) or dominant-negative AMPK as reported (10).

Immunoprecipitation and Western Blotting

For immunoprecipitation human endothelial cells were lysed in Triton X-100, left on ice for 10 min, and centrifuged at 10,000 g for 10 min. After being precleared with protein A/G Sepharose, proteins were immunoprecipitated from the cell supemate with phosphotyroline antibody (Santa Cruz Biotechnology, Heidelberg, Germany). For Western blot analysis, cells were lysed in Triton X-100 lysis buffer and separated by SDS-PAGE as described (30).

Reporter Gene Assay

Endothelial cells overexpressing either VEGFR1 or VEGFR2 were transiently cotransfected with the noncoding 5′ region of CYP2C9 (−2,088 to +21; kindly provided by Dr. P. Maurel, Montpellier, France), subcloned into pGL3basic (Promega, Mannheim, Germany) and a plasmid containing lacZ under the control of the cytomegalovirus (CMV) promoter. After 12 h, the cells were treated with either solvent (PBS, 0.6%) or VEGF (30 ng/ml) for 6 h. Thereafter, luciferase (Promega, Mannheim, Germany) and β-galactosidase (Tropix, Bedford, MA) activity was assayed according to the manufacturers' protocols. Promoter activity was quantified as luciferase activity relative to that of β-galactosidase and normalized to the respective nontreated controls.

Isolation of RNA and RT-qPCR

Total RNA was isolated from cultured endothelial cells using phenol and guanidine isothiocyanate (TriReagenz, Sigma, Germany). Random hexanucleotide primers were used for reverse transcription of equal amounts of RNA. The cDNA was used for real-time PCR using Taqman probes for the detection of the specific amplification products. For the detection of CYP2C8 mRNA by real-time RT-PCR the oligonucleotides were used as the following: 2C8 forward: 5′-GGAGTTATCGATGTCCTGG-3′, reverse: 5′-CCTATGTAGAGTGCTGGCTTG-3′; FAM- and dabycyl-labeled taqman probe: 5′-TACACGTATCCTCTTTGAATGGCTCCTAT-3′ corresponding to base pair positions 4301 to 4319 (forward), 4550 to 4567 (reverse) and 4381 to 4404 in RNA Pol2 cDNA (NCBI accession number NM_000937). The amount of cDNA in the samples was calculated on the basis of the amplification of a serial dilution of a plasmid (CYP2C8) or the serial dilution of the cDNA (RNA Pol2). The CYP2C8 levels were normalized to that of Pol2. At least two RT reactions were performed using each RNA preparation and at least two PCR reactions were performed with each cDNA sample.

Antisense Oligonucleotides

In some experiments, an antisense oligonucleotide approach was used to prevent the VEGF-induced upregulation of CYP2C, as described (32). Cells were treated with CYP2C sense and antisense oligonucleotides (2 μmol/l; antisense: 5′-TCCATTGGAAGCTTCTCTTCTTCT-3′; sense: 5′-AAGAAGAGAGGCTTACAGGA-3′; in both cases the three 5′ nucleotides were modified with phosphothioate; MWG-Biotech, Ebersberg, Germany) according to manufacturer’s protocol (MobiTec, Göttingen, Germany). The sequence of these oligonucleotides spans the ATG and is 100% identical with human CYP2C8 and contains one mismatch to the other three human CYP2C isoforms.

Regulation (siRNA) of the AMPK

To downregulate the AMPK α-subunit in human and murine endothelial cells specific small interfering RNAs (siRNAs) were generated (Eurogentec, Searing, Belgium). The sequence used to target the human isoform was CCAAGUGGUAGUAGAACCU-dTdT, whereas a mixture or two siRNAs were used to target the murine isoform (GGACCAUCUUAAUGUACA-dTdT and GGUCAUCAGACACACAU-dTdT; only the coding strand of the duplex is indicated). For siRNA transfection, Gene trans II was used according to the manufacturer’s protocol. For the Matrigel plug assay, equal amounts (2 μmol/l) of both murine siRNAs were added to Matrigel just before implantation (see below).

Liquid Chromatography-Mass Spectrometry Measurements

Human endothelial cells, treated as described in RESULTS and harvested by scraping, and the pellets (from ~8 × 10^6 cells) were suspended in 100 μl potassium phosphate buffer (0.1 mol/l, pH 7.2), hydrolyzed for 1 h in NaOH (0.5 N), and neutralized with HCl (2 mol/l) before deuterated internal standards (5-HETE-d8, 12-HETE-d8, 15-HETE-d8, 20-HETE-d6, 8,9-EET-d8, 11,12-EET-d8, and 14,15-EET-d8) were added. A liquid-liquid extraction was performed twice using ethyl acetate (0.5 ml). After evaporation of the solvent in a vacuum block under a gentle stream of nitrogen, samples were reconstituted with 50 μl of methanol-water (1:1, vol/vol) and eicosanoids were determined with a Sciex API4000 mass spectrometer operating in the multiple reaction monitoring mode. Chromatographic separation was performed on a Gemini C18 column (150 × 2 mm ID, 5-μm particle size, Phenomenex, Aschaffenburg, Germany).

In Vitro Angiogenesis Assays

Tube formation. Primary cultures of endothelial cells were seeded on cell culture dishes coated with fibronectin and cultured in MCDB 133 medium containing 4% FCS and either solvent or VEGF (30 ng/ml) for 6 h. Thereafter, cell culture dishes were seeded with endothelial cell spheroids. Spheroids containing 400 cells were reconstituted with 50 μl of methanol-water (1:1, vol/vol) and eicosanoids were determined with a Sciex API4000 mass spectrometer operating in the multiple reaction monitoring mode. Chromatographic separation was performed on a Gemini C18 column (150 × 2 mm ID, 5-μm particle size, Phenomenex, Aschaffenburg, Germany).
Matrigel plug assay. Female C57BL/6 mice (8 wk old) were lightly anesthetized with chloral hydrate (200 μl of a 4% solution injected subcutaneously) and then injected subcutaneously with 0.5 ml of Matrigel impregnated with heparin (0.0025 U/ml), VEGF (150 ng/ml), bFGF (150 ng/ml), EEZE (100 μmol/l), or combinations thereof, along the dorsal midline on each site of the spine. Fourteen days later the mice were euthanized, the Matrigel plugs were removed, embedded in tissue tech (Sakura Finetec), and frozen. Plugs were sectioned (10 μm) by cryosection and processed for staining for CD31 (BD Biosciences, San Jose, CA) or α-smooth muscle actin (Sigma). Afterwards preparations were mounted and viewed using a confocal microscope (LSM 510 META, Zeiss). Vessel formation was quantified by analyzing at least five five sections per plug. The investigation conforms with the Guide for the Care and Use of Laboratory Animals published by the National Institutes of Health (NIH Publication No. 85-23, Revised 1996), assurance number A5626-01. The angiogenic response was graded by three observers who were blind to the experimental conditions. The vessel formation index scores were as follows: 0, no effect, no invaded cells; 1, few invaded endothelial cells; 2, clear invasion of endothelial cells; 3, clear invasion of endothelial cells and capillary formation; and 4, endothelial and smooth muscle cell invasion, clear vessel formation. To facilitate comparison between the different groups the scores were normalized with respect to the effects observed in the control group.

Statistics

Data are expressed as means ± SE. Statistical analysis of Matrigel plugs analyzed by ultrasound was performed using a paired t-test. For all other statistical comparisons evaluation was performed with Student’s t-test for unpaired data or one-way ANOVA followed by a Bonferroni t-test. Values of P < 0.05 were considered statistically significant.

RESULTS

Effect of VEGF on CYP2C Expression and Activity

To assess the effects of VEGF on CYP2C expression we measured the activity of a reporter construct as well as levels of endogenous CYP2C RNA and protein. VEGF treatment (30 ng/ml, 6 h) of porcine aortic endothelial cells overexpressing either VEGFR1 or VEGFR2 and transfected with a CYP2C promoter-luciferase construct elicited a significant increase in the activity of the reporter gene construct (Fig. 1A). The increase in promoter activity in the cell line was paralleled by a VEGF-induced increase in CYP2C8 mRNA (Fig. 1B) in primary cultures of human endothelial cells.

As reported previously, although CYP2C protein can easily be detected in native endothelial cells, its expression decreases rapidly following cell isolation (12, 18). Indeed, only low levels of CYP2C protein were detectable in the human umbilical vein endothelial cells studied (Fig. 2A). Simulation of the endothelial cells with VEGF, however, induced the expression of CYP2C (Fig. 2A) as well as CYP epoxygenase activity and the generation of 11,12-EET (Fig. 2B), which was sensitive to the epoxygenase inhibitor, miconazole.

Role of CYP in VEGF-Induced Endothelial Cell Tube Formation

To assess the involvement of CYP epoxygenases in VEGF-induced endothelial cell tube formation, human umbilical vein endothelial cells were seeded onto fibronectin-coated culture dishes and treated with VEGF (30 ng/ml, 48 h) in the absence and presence of the EET antagonist 14,15-EEZE (10 μmol/l) or the CYP epoxygenase inhibitor MS-PPOH (10 μmol/l). Whereas endothelial cell tubes were apparent after 48 h in the VEGF-treated cultures, the number of tubes formed was significantly attenuated in cells treated with either the EET antagonist or the CYP inhibitor (Fig. 3A). To determine whether the effects observed were specific to VEGF-activated signaling, we repeated the experiments using bFGF as an angiogenic stimulus. In the latter case, interfering with CYP generation and EET activity was without effect on the ability of the growth factor to stimulate tube formation (Fig. 3A).

To ensure that the inhibitory effects observed using EEZE did not result from any direct interference between the EET antagonist and VEGF signaling, we assessed its effect on the VEGF-induced phosphorylation of VEGFR2. In VEGFR2 expressing endothelial cells, there was a low but detectable basal phosphorylation of the receptor, which was increased significantly by the application of VEGF (30 ng/ml, 20 min). 14,15-EEZE failed to affect the tyrosine phosphorylation of VEGFR2 under basal conditions or following stimulation with VEGF (Fig. 3B).

To more directly assess the impact of CYP2C on VEGF-induced endothelial cell sprouting in vitro, we performed a collagen-based spheroid assay and combined it with a previously described antisense oligonucleotide approach (13) to prevent the VEGF-induced increase in CYP2C expression. VEGF (30 ng/ml, 24 h) stimulated endothelial cell

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Fig. 1. Effect of vascular endothelial growth factor (VEGF) on the cytochrome P-450 (CYP) CYP2C gene in endothelial cells. A: CYP2C9 promoter activity in porcine endothelial cells overexpressing VEGFR1 or VEGFR2 were transfected with the promoter construct 14 h before stimulation with VEGF (30 ng/ml, 6 h). B: time course showing the effect of VEGF (30 ng/ml, 3–48 h) on the expression of CYP2C mRNA in human umbilical vein endothelial cells. Bar graphs summarize data obtained in 7–13 independent experiments. *P < 0.05, **P < 0.001 vs. control (CTL).
sprouting (Fig. 4); however, transfecting endothelial cells with CYP2C antisense oligonucleotides 14 h before generating the spheroids markedly attenuated VEGF-induced sprouting. CYP2C sense oligonucleotides used as a control were without effect on the response to VEGF (Fig. 4).

Similarly, CYP2C antisense oligonucleotides also prevented VEGF-induced tube formation (Fig. 4).

Role of the AMPK in the VEGF-Induced Induction of CYP2C

The AMPK has been implicated in both VEGF-induced angiogenesis (33) as well as the induction of CYP expression by phenobarbital (37). We therefore determined the involvement of the AMPK in the VEGF-induced upregulation of CYP2C expression.

VEGF induced a rapid (within 5 min) phosphorylation of the AMPK on Thr172. The response was biphasic with an initial peak at 10 min (813 ± 55% of control, n = 4, P < 0.001) followed by a return to basal levels and a secondary but sustained AMPK activation after 24 h (593 ± 120% of control, n = 4, P < 0.01; Fig. 5A). The overexpression of constitutively active AMPKα increased CYP2C RNA levels in endothelial cells, whereas the overexpression of a dominant negative AMPKα mutant prevented the VEGF-induced increase in CYP2C RNA (Fig. 5B). The dominant negative AMPKα mutant also prevented the VEGF-induced increase in CYP2C protein (Fig. 5C) as well as endothelial cell tube formation (Fig. 5D). The CYP inducer phenobarbital (1 mmol/l) stimulated the phosphorylation of AMPKα in endothelial cells (Fig. 5A) and increased endothelial CYP2C expression (Fig. 5C).

Effect of 14,15-EEZE on VEGF-Induced Angiogenesis In Vivo

To assess the role of CYP-derived EETs on VEGF-induced angiogenesis in vivo, we determined the endothelial cell content of Matrigel plugs that were impregnated with either solvent, VEGF (150 ng/ml), the EET antagonist 14,15-EEZE...
cell invasion of the plugs, this effect was insensitive to the EET antagonist (Fig. 6B).

**Effect of AMPK Downregulation on VEGF-Induced Angiogenesis**

Because our data indicated that the VEGF-induced increase in CYP2C expression is dependent on the activation of the AMPK, we next assessed the consequences of AMPK downregulation on endothelial cell tube formation in vitro in human endothelial cells and endothelial cell invasion of Matrigel in vivo in mice.

The siRNA used almost abrogated AMPKα expression in human endothelial cells (Fig. 7A) and prevented the VEGF-induced formation of tube-like structures (Fig. 7B). The same procedure was repeated using murine lung endothelial cells in which we compared the ability of two siRNAs to downregulate the AMPK (Fig. 7C). Both of the siRNA tested downregulated the AMPK with one of them (siRNA2) being more effective.

To determine whether the AMPK siRNA could also affect angiogenesis in vivo, Matrigel plugs were then impregnated with either solvent or VEGF together with either a control siRNA (GFP) or a mixture of the AMPK siRNA1 and siRNA2 and implanted in mice for a total of 7 days. Little or no PECAM-1 was detected in plugs impregnated with solvent (data not shown). Moreover, whereas PECAM-1-positive endothelial cells could be detected in plugs treated with VEGF and the control siRNA, PECAM-1 staining was sparse in plugs treated with VEGF and the AMPK siRNA (Fig. 7D).

**DISCUSSION**

The results of the present investigation demonstrate that stimulation of endothelial cells with VEGF results in an increase of CYP2C protein as well as of 11,12-EET levels. This activation of CYP2C and subsequent EET release seems to be dependent on the AMPK and to be an essential component of the signaling cascade initiated by VEGF as endothelial cell sprouting was abolished by CYP2C downregulation (antisense oligonucleotides). In addition, the CYP epoxygenase inhibitor MS-PPOH and EET antagonist 14,15-EEZE selectively inhibited VEGF-, but not FGF-mediated, angiogenesis.

Although interest in the CYP epoxygenases expressed in the vasculature was focused on the role played by the EETs in the regulation of membrane potential, these eicosanoids are now recognized as signaling molecules, able to stimulate several steps in the angiogenic process, including endothelial cell proliferation, migration, and tube formation (31). However, the majority of the studies performed to date have relied on overexpression systems, and little is known about the involvement of CYP epoxygenases/EETs in angiogenic pathways stimulated by classical growth factors. Because we recently reported that EETs are involved in hypoxia-induced angiogenesis (29), it seemed logical to hypothesize a potential link between VEGF and EET signaling, since the induction of VEGF by hypoxia and the role of the latter growth factor in hypoxia-induced cell migration is well documented (11). The results of our study demonstrate that endothelial cell stimulation with VEGF increases CYP2C expression and EET production and that preventing the induction or inhibiting the activity of CYP2C significantly attenuates the
angiogenic response to VEGF. These observations fit well with a recent report indicating that another CYP-derived metabolite of arachidonic acid, namely 20-HETE, can be activated by VEGF in pulmonary arteries (23).

The CYP2C enzyme sensitive to VEGF was identified as CYP2C8, and we were able to show that both CYP2C8 mRNA and protein increased in the cells studied. One limitation of the present study was that we were only able to analyze the effects of VEGF on the activity of a CYP2C9 and not a CYP2C8 promoter construct. However, the enzymes are highly homologous and both have been implicated in angiogenesis. Moreover, we have frequently found similar effects (Ca²⁺ signaling and angiogenesis) in native endothelial cells, which express CYP2C8 and in cultured endothelial cells overexpressing CYP2C9 (18, 41). The stereochemistry of the EETs produced by these two enzymes is however quite different, thus it will be important to determine whether different effects in angiogenesis and cell signaling in general are observed using different EET enantiomers. Which of the EETs is responsible for the effects reported also remains to be clarified, and although we have repeatedly been able to reproduce the effects of CYP activation/overexpressing using 11,12-EET and not 14,15-EET (30, 41), this finding is not shared by others (28).

The importance of VEGF and its receptors in vascular development has been demonstrated in various knockout animals. For example, the deletion of either VEGFR1 (38) or VEGFR2 (19) has proven to be lethal due to abnormalities in vessel formation, therefore, it appears both receptors are essential for the development of the vasculature in the mouse embryo. Analysis of VEGFR signaling has led to the conclusion that although affinity for VEGF binding is approximately 10-fold higher for VEGFR1 than for VEGFR2, it is activation of the latter that is believed to convey the VEGF-mediated effects in endothelial cells (21). We observed however that VEGF was able to increase the activity of the CYP2C9 promoter in cells expressing either VEGFR1 or VEGFR2. Although EETs have been suggested to act as second messengers in the EGF-activated signaling cascade (5), not all growth factors are able to upregulate CYP epoxygenases, and we found that although the EET antagonist 14,15-EEZE almost abolished VEGF-induced endothelial tube formation in vitro and in vivo, it did not affect the angiogenesis induced by bFGF.

In the search to identify the molecular mechanism(s) underlying the VEGF-induced increase in CYP2C expression, we analyzed the role of the AMPK as VEGF has previously been demonstrated to activate the kinase in endothelial cells (36), and activation of AMPK has been linked to endothelial cell migration and angiogenesis (33, 34). Moreover, activation of the AMPK by phenobarbital is reported to increase the expression of CYP2B2 in chicken and human hepatocytes (1, 37). Consistent with these reports, both VEGF and phenobarbital stimulated the phosphorylation of the AMPK and increased in CYP2C expression in endothelial cells. Moreover, the overexpression of a dominant-negative AMPK mutant significantly reduced VEGF-induced CYP2C expression, and AMPK downregulation markedly attenuated VEGF-induced angiogenesis in vitro and in vivo. Although we found this involvement of AMPK in the VEGF-induced CYP2C expression, the remaining steps that involved in this process have not been elucidated, but LKB1 (serine/threonine kinase 11) is likely to be involved (1).

The VEGF-activated signaling cascade leading to angiogenesis has previously been linked to an increase in the formation of reactive oxygen species (9). Although the NADPH oxidase...
Fig. 6. Effect of 14,15-EEZE on VEGF- and bFGF-induced angiogenesis in vivo. A: endothelial cell migration into Matrigel plugs impregnated with either solvent (DMSO, 0.1%, CTL), VEGF (150 ng/ml), the EET antagonist 14,15-EEZE (100 μmol/l), or the combination of both VEGF and EEZE. Images were taken after sectioning and immunostaining for CD31 (red) and β-actin (green).

B: endothelial cell migration into Matrigel plugs impregnated with either solvent (Sol), bFGF (150 ng/ml), the EET antagonist 14,15-EEZE (100 μmol/l), or the combination of both bFGF and EEZE. Images were taken after sectioning and immunostaining for CD31 (red) and β-actin (green) and scored as the vessel formation index (see METHODS). Bar graphs summarize data obtained in 4–10 independent experiments. *P < 0.05, ***P < 0.001 vs. the appropriate solvent (Sol) in the absence of EEZE, §§§P < 0.001 vs. VEGF.
has been implicated as the source of these radicals (40), it should be noted that the activation of CYP2C epoxygenases also results in the generation of superoxide anions ($\text{O}_2^{-}/\text{H}_2\text{O}_2$) in sufficient amounts to alter the bioavailability of nitric oxide and the expression of adhesion molecules (17). To verify that CYP2C-derived EETs rather than reactive oxygen species were involved in mediating VEGF-induced angiogenesis, we employed the EET antagonist 14,15-EEZE. In contrast to the epoxygenase inhibitors MS-PPOH and sulfaphenazole, as well as the CYP2C antisense treatment, which attenuate the production of all the CYP2C products (EETs, 20-HETE, and $\text{O}_2^{-}/\text{H}_2\text{O}_2$), 14,15-EEZE antagonizes only the effects of EETs without interfering with those of 20-HETE (20) or the production of $\text{O}_2^{-}$ by CYP2C enzymes (U. R. Michaelis, unpublished observations). We found that 14,15-EEZE inhibited VEGF-induced tube formation in vitro and angiogenesis in vivo, indicating that EETs and not reactive oxygen species were responsible for the effects observed. Moreover, we were unable to detect an effect of VEGF on 20-HETE formation, indicating that 20-HETE does not contribute to angiogenesis under the conditions studied.

Taken together our data indicate the potential importance of 11,12-EET to (patho)physiological angiogenic processes. At this point it is important to point out that CYP enzymes are not only expressed in endothelial cells and CYP2C as well as CYP2J enzymes have been detected in different tumor tissues (24, 42) and can induce tumor growth as well as promote metastasis (25). It is therefore tempting to speculate that CYP enzymes might represent a new target for the treatment of tumor growth and therefore cancer therapy. Pharmacological inhibitors of some CYP isoforms have been identified as promising anti-cancer agents (7, 8); however, the majority of work published to date has focused on the consequences of CYP inhibition on the bioavailability of anti-cancer agents rather than determining the consequences of CYP inhibition per se.

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