The role of mitochondria in pharmacotoxicology: a reevaluation of an old, newly emerging topic

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Scatena R, Bottoni P, Botta G, Martorana GE, Giardina B. The role of mitochondria in pharmacotoxicology: a reevaluation of an old, newly emerging topic. Am J Physiol Cell Physiol 293: C12–C21, 2007. First published May 2, 2007; doi:10.1152/ajpcell.00314.2006.—In addition to their well-known critical role in energy metabolism, mitochondria are now recognized as the location where various catabolic and anabolic processes, calcium fluxes, various oxygen-nitrogen reactive species, and other signal transduction pathways interact to maintain cell homeostasis and to mediate cellular responses to different stimuli. It is important to consider how pharmacological agents affect mitochondrial biochemistry, not only because of toxicological concerns but also because of potential therapeutic applications. Several potential targets could be envisaged at the mitochondrial level that may underlie the toxic effects of some drugs. Recently, antiviral nucleoside analogs have displayed mitochondrial toxicity through the inhibition of DNA polymerase-γ (pol-γ). Other drugs that target different components of mitochondrial channels can disrupt ion homeostasis or interfere with the mitochondrial permeability transition pore. Many known inhibitors of the mitochondrial electron transfer chain act by interfering with one or more of the respiratory chain complexes. Nonsteroidal anti-inflammatory drugs (NSAIDs), for example, may behave as oxidative phosphorylation uncouplers. The mitochondrial toxicity of other drugs seems to depend on free radical production, although the mechanisms have not yet been clarified. Meanwhile, drugs targeting mitochondria have been used to treat mitochondrial dysfunctions. Importantly, drugs that target the mitochondria of cancer cells have been developed recently; such drugs can trigger apoptosis or necrosis of the cancer cells. Thus the aim of this review is to highlight the role of mitochondria in pharmacotoxicology, and to describe whenever possible the main molecular mechanisms underlying unwanted and/or therapeutic effects.

mitochondrial diseases; nitric oxide; apoptosis; degenerative diseases; free radicals

MITOXONDRIA CAN REPRESENT a primary or secondary drug target (102, 112). However, some aspects of drug-mitochondria interactions may still be underestimated because of the difficulty in foreseeing and understanding all potential implications of the complex pathophysiology of mitochondria. Insufficient consideration of mitochondrial pharmacotoxicology may also be due to a lack of knowledge about acquired mitochondrial diseases, which are a heterogeneous and growing class of disorders ranging from Type 2 diabetes to neurodegenerative diseases and cancer (30, 32, 105).

A pathogenetic role for mitochondrial dysfunction has been invoked in a vast number of illnesses without sufficient experimental support to precisely establish the molecular pathophysiological mechanisms. Indeed, mitochondrial physiology and pathophysiology are very complex, and the role of the organelles in bioenergetics is strictly linked to other essential functions such as anabolic pathways, redox balance, cell death and differentiation, and mitosis, along with more specialized cell functions including calcium homeostasis and thermogenesis, reactive oxygen species (ROS) and reactive nitric oxide species signaling, ion channels, and metabolite transporters. The same complexity and heterogeneity can be surmised from the range of congenital mitochondrial diseases, providing further evidence of the difficulty in correctly approaching mitochondrial pathophysiology.

These unique aspects of mitochondria should stimulate us to pay more attention than that usually devoted to both the toxic and therapeutic aspects of the interrelationships between drugs and mitochondria. Moreover, their typical structural and functional characteristics may make mitochondria a valuable target for xenobiotics (30, 32).

Mitochondria: Structure-Function and Pharmacotoxicology Relationships

Two main structural and functional aspects of mitochondria should first be considered: the presence of different organelle subcompartments in mitochondria and the fact that mitochondria have their own DNA. Structurally, mitochondria are very diverse across organs and tissues, but all mitochondria contain two lipid bilayer membranes. The outer membrane delineates the organelle and is structurally similar to other cell membranes, being rich in cholesterol and permeable to ions. The
inner membrane, which isolates the matrix, is virtually devoid of cholesterol, is rich in cardiolipin (which binds the proteins of the electron transport chain), and is impermeable to ions. This impermeability accounts for the generation of the electrochemical gradient that supplies the proton motive force for ATP generation. Therefore, the maintenance of the integrity of this inner mitochondrial membrane is critical for mitochondrial function. This typical structure is suitably organized to perform and finely coordinate all of the distinctive activities of mitochondria. In fact, in addition to their critical role in energy generation in eukaryotic cells, mitochondria are also active participants in a variety of tissue-specific metabolic processes, such as urea generation, heme synthesis, and fatty acid β-oxidation. Mitochondria are able to carry out all of these functions because they are characterized by a unique milieu, with an alkaline and negatively charged interior and a series of specific channels and carrier proteins. Importantly, the complex structure and typical physicochemical characteristics [mainly mitochondrial membrane potential (ΔΨ) and pH = 8] facilitate the selective accumulation of xenobiotics in the matrix and/or the inner mitochondrial membrane by exerting an efficient trap effect (79, 100). As a consequence, mitochondria can easily accumulate lipophilic compounds of cationic character and, even better, weak acids in their anionic forms. Importantly, for the latter in particular, their undissociated forms can penetrate the inner mitochondrial membrane freely, while their protons dissociate when inside the alkaline matrix, rendering the molecules much less permeant and trapping them inside the organelle (7). Such mitochondrial drug storage may interfere with the determination of pharmacokinetic parameters (distribution volume, plasma concentration, and half-life). Moreover, a vicious circle could then ensue, possibly leading to the progressive accumulation of these acid lipophilic xenobiotics inside mitochondria, damaging their function and permeability properties. Depending on the level and number of mitochondria affected, the cell could go toward a grave deenergization state that in turn can lead to necrosis or more localized damage of a few mitochondria with a collapse of their ΔΨ. The resultant pH modification could reprotox the xenobiotic, rendering it freely permeant in the cell and capable of entering other mitochondria (91, 92). This process may thus allow a progressive spread of the xenobiotic.

Mitochondria are the only organelles outside of the nucleus that contain DNA. The mitochondrial genome consists of a small circular chromosome that contains a total of 37 genes. Thirteen of these encode proteins that are unique components of the electron transport chain. The remaining genes encode 22 tRNAs and 2 ribosomal RNAs used in the mitochondrial ribosome subunits. The result is that the mitochondrion is fully capable of synthesizing at least some proteins (the remaining proteins are the products of nuclear genes and are synthesized in the cytosol and translocated into the mitochondria). This capability is essential to its function in energy generation, but exposes it to unique risks.

Unlike nuclear DNA, mitochondrial DNA (mtDNA) is not protected by histone proteins. Furthermore, mitochondria are located in close proximity to sites where ROS are routinely generated. However, DNA repair processes are generally less efficient for mtDNA. As a result, mtDNA is more likely to undergo mutation than nuclear DNA; the mutation rate is estimated to be at least 10–20 times higher (9).

mtDNA can undergo replication and, to a limited extent, base excision repair. Both of these functions reside in a single DNA polymerase, polymerase-γ (pol-γ), in mitochondria (in contrast to nuclear DNA, which is maintained by at least 9 polymerases). Although pol-γ is a nuclear protein, it has no known function other than mtDNA replication, and thus any mutation or inhibition of this enzyme will be manifested only in mtDNA.

All of this underscores the fact that mitochondria are potential primary or secondary targets of xenobiotics and that the interaction of xenobiotics with mitochondria or mitochondrial components (i.e., mtDNA, respiratory chain complexes, biomembranes with their different transporters, or matrix metabolic enzymes) should not always be considered negative. In fact, recent data show that an unexpected therapeutic effect could result from pharmacological modulation of the organelle’s activities (30, 32, 79, 91, 92). Many chemicals are known to interact with mitochondrial molecules (22, 77, 83); however, an in-depth discussion of all the known interactions is beyond the scope of this article and would not be possible because of space constraints. The aim of this review is to discuss the role of mitochondria in general and of their subcompartments in particular in pharmacotoxicology, describing whenever possible the main molecular mechanisms underlying unwanted and therapeutic effects. Such knowledge could stimulate interest in the possible incidence of iatrogenic mitochondrialopathies and, moreover, promote a real mitochondrial pharmacology with potential therapeutic applications in a growing number of prominent disease states, including ischemia-reperfusion injury, neurodegenerative diseases, cancer, and hyperlipidemias.

**Mitochondria and Drugs: Toxicological Issues**

Mitochondria play a critical role in supplying the cell with the bulk of its ATP needs via oxidative phosphorylation (oxphos); thus any cell type or tissue with a high aerobic energy requirement is more likely to be affected when this organelle is dysfunctional. In addition, the enzymes necessary for several specialized metabolic processes (fatty acid β-oxidation, urea synthesis, heme synthesis) reside within the mitochondrial matrix. As a result, tissues that rely heavily on these processes are also frequent targets of mitochondrial toxins. For these reasons there are numerous common syndromes associated with mitochondrial toxicity including lactic acidosis, cardiac and skeletal myopathy, peripheral, central, and optic neuropathy, retinopathy, ototoxicity, enteropathy, pancreatitis, diabetes, hepatic steatosis, and hemotoxicity. Combinations of these effects (or different manifestations of toxicity in different individuals treated with the same compound) are not uncommon and are strong indicators that the underlying toxic insult involves mitochondria. Mitochondrial toxicities in general tend to be chronic injuries with somewhat variable manifestations. Most cells contain a large number of mitochondria that allow for some functional reserve, and cellular injury or dysfunction will occur only when enough mitochondria are irreparably damaged and the cell cannot meet its energy demands. When cells divide, the mitochondria apportionment between them is random ("heteroplasmy"): one daughter cell may contain primarily normal mitochondria while the other gets a dispropor-
tionate share of damaged mitochondria, resulting in a patchy distribution of damaged cells within a tissue.

**Mitochondrial DNA.** mtDNA may be damaged by drugs through different mechanisms. A well-known exogenous agent capable of oxidatively damaging mtDNA is ethanol (26). Some drugs can selectively damage mtDNA by inhibiting its synthesis, as has been recently exploited with the introduction of nucleotide reverse transcriptase inhibitors (NRTIs). These compounds are nucleoside analogs that are taken up by cells and sequentially phosphorylated to the active triphosphate form (22, 66). The nucleotide triphosphates can thus be used as substrates by retroviral reverse transcriptase, while their incorporation into the nascent DNA chain results in chain termination. The triphosphates of these analogs have also been shown to be potential substrates for pol-γ, the unique mtDNA polymerase, and can similarly result in chain termination during mtDNA replication (22). Additional effects on mtDNA synthesis result from the fact that the conversion of the monophosphorylated to the triphosphorylated form is extremely inefficient within mitochondria. Consequently, these monophosphorylated forms can build up to high (mM) levels in the mitochondrial matrix and at such high levels can have other effects on mtDNA synthesis. These include inhibition of the exonuclease function of pol-γ (resulting in decreased replication fidelity) and also, as recently shown with zidovudine, may significantly inhibit thymidine phosphorylation, thus affecting DNA replication by depletion of a necessary substrate (111). This DNA pol-γ dysfunction, which induces a progressive depletion of mtDNA, ultimately interferes with the synthesis of essential proteins of the mitochondrial respiratory chain (MRC) (65, 72). The consequent disruption of the electron respiratory chain results in reduced ATP synthesis and electron leakage, leading to increased production of free radical species. Enzyme assay and cell culture studies of NRTIs have demonstrated the following hierarchy of mtDNA pol-γ inhibition: zalcatibine > didanosine > stavudine > lamivudine > zidovudine > abacavir (96). In vitro investigations have also documented impairment of mitochondrial adenylate kinase and the adenosine diphosphate/adenosine triphosphate translocator. Inhibition of pol-γ and other mitochondrial enzymes can gradually lead to critical mitochondrial dysfunction and cytotoxicity. The clinical manifestations of NRTI-induced mitochondrial toxicity resemble those of inherited mitochondrial diseases, i.e., hepatic steatosis, lactic acidosis, myopathy, peripheral neuropathy, and, intriguingly, nephrotoxicity. Fat redistribution syndrome, or human immunodeficiency virus (HIV)-associated lipodystrophy, is another side effect attributed to NRTI therapy (18). The morphological and metabolic complications of this syndrome are similar to those of the mitochondrial disorder known as multiple symmetric lipomatosis, suggesting that this too may be related to mitochondrial toxicity (59, 65, 117).

**Mitochondrial respiratory chain.** Drug-induced derangements of the MRC can occur at any of the four protein complexes in the respiratory chain. Effects on complex IV (cytochrome-c oxidase), however, are the most severe because this is the step where oxygen is reduced to water. Inhibition of complex III can also frequently result in the generation of ROS as a consequence of the intrinsic characteristics of the electron transfer process to this complex from reduced ubiquinone.

With respect to iatrogenic mitochondrialopathies, many molecules are well-known inhibitors of mitochondrial complexes. Some of these toxic compounds (including rotenone, antimycin, cyanide, oligomycin, and mixtothiazol) have been widely employed to analyze the function of the mitochondrial bioenergetic machinery in general and of electron transport in particular, and have already been the subject of numerous exhaustive reviews and books (33, 40, 50, 52). Interactions of drugs of clinical interest for the mitochondrial electron respiratory chain have not been as well studied. Nevertheless, several drugs are known to act by partially inhibiting, directly and/or indirectly through their metabolites, components of the MRC; examples of such drugs include amiodarone, perhexiline, flutamide, and anthralin (26, 35, 42, 43). The molecular mechanisms by which these drugs impair the MRC have not been resolved. Two hypotheses have been postulated: 1) a direct inhibition of a protein subunit of one or more enzyme complexes and 2) an electron diversion from the MRC by drugs that act as spurious acceptors (60).

Strong support for the importance of these iatrogenic mitochondrialopathies has been provided by a chemically induced parkinsonism resulting from accidental poisoning from 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). The condition is produced as an impurity in batches of the illegally made “synthetic heroin” 1-methyl-4-phenyl-1-propylnorpyriderine (MPPP). In particular, this by-product, via its metabolite MPP⁺, seems to mainly affect complex I of the mitochondrial oxphos pathway preferentially in dopaminergic neurons (61). Importantly, experimental and clinical data showed that such iatrogenic parkinsonism is characterized by the same neuropathological features as the idiopathic forms (i.e., loss of substantia nigra dopaminergic neurons, Lewy bodies, and typical accumulation of neuromelanin in microglia and extracellular matrix) (34).

There are other molecules that are usually considered real complex I inhibitors, including some well-established drugs (papaverine, meperidine, cinnarzine, amytal, haloperidol, and ketoconazole and its analogs) (38, 78, 98, 101, 109). These molecules share a common structural motif, a cyclic head and a hydrophobic tail (25, 31, 32, 74). Intriguingly, this typical structure is also present in another class of therapeutic agents, the so-called fibrates (clofibrate acid, bezafibrate, gemfibrozil) and in some of their derivatives, the thiazolidinediones (ciglitazone, troglitazone, pioglitazone). Our studies (90, 92), recently confirmed by Brunmair et al. (11, 12), showed that these compounds can also inhibit mitochondrial complex I, resulting in the metabolic consequences typical of their pharmacological activities (hypolipidemic and hypoglycemic effects); such inhibition may explain some of their toxic effects (rhabdomyolysis, acute liver failure) that intriguingly resemble those of inherited mitochondrialopathies. More than 60 types of compounds are well-known inhibitors of complex I, and this number continues to grow. This tendency of xenobiotics to inhibit complex I (mitochondrial NADH:ubiquinone oxidoreductase; EC 1.6.5.3) may depend on the intricate structure of this enzymatic complex, which consists of at least 40 different polypeptides strongly embedded in the inner mitochondrial membrane. This unique feature explains the mitochondrion’s great vulnerability to lipophilic molecules (25, 30, 74). Regarding potential toxicity, complex I is a secondary target of nitric oxide (NO) in general and of nitrogen radical
species in particular; this should be kept in mind when patients are being treated with old and particularly new NO donor drugs (10, 15, 76, 85, 89).

Complex II of the electron respiratory chain, succinate dehydrogenase (SDH), is less commonly studied in mitochondrial pharmacotoxicology, which is surprising considering that it also plays a role in the tricarboxylic acid cycle. Apart from the common inhibitors usually employed in experimental studies (i.e., malonate, carboxin, 3-nitropropionic acid), it is worth pointing out that some cis-crotonalide fungicides, diazoxide, and, more recently, some fluoroquinolones, chloramphenicol succinate, and anthracycline drugs are complex II inhibitors that also inhibit other mitochondrial components (102, 112). Importantly, recent reports regarding the role of SDH (or of one of its components: the B, C, and D subunits) in tumor susceptibility have opened up a new perspective in research on the modulation of oncosuppression and/or oncopromotion by mitochondria (4, 8, 45, 83, 95). The mechanism of this tumor promotion by SDH and by fumarate hydratase (FH) has been ascribed to an intriguing metabolic signaling pathway that starts with the physiological substrates of these enzymes (i.e., succinate and fumarate, respectively). In fact, these metabolites accumulate in mitochondria because of the inactivation and/or low activity of SDH and FH, leak out to the cytosol, and there inhibit a family of prolyl hydroxylase enzymes. This inhibition, in turn, may render neoplastic cells more resistant to apoptotic signals and activate a pseudohypoxic response (mediated by hypoxia-inducible factor) that enhances glycolysis (84). Other signals and activate a pseudohypoxic response (mediated by hypoxia-inducible factor) that enhances glycolysis (84). Other signals and activate a pseudohypoxic response (mediated by hypoxia-inducible factor) that enhances glycolysis (84). Other signals and activate a pseudohypoxic response (mediated by hypoxia-inducible factor) that enhances glycolysis (84). Other signals and activate a pseudohypoxic response (mediated by hypoxia-inducible factor) that enhances glycolysis (84).

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effectors responsible for cellular and/or molecular damage. There are also xenobiotics that are capable of inducing ROS generation without directly deranging the MRC, examples of which are haloalkenyl cysteine conjugates such as hexachlorobutadiene (which form reactive thiols subsequent to their activation by the mitochondrial enzyme β-lyase), 4-thiaalkanones (activated by fatty acid β-oxidase), and valproic acid (activated by acyl-CoA synthase) (39, 106).

Oxidative phosphorylation. Compounds that dissipate the proton gradient between the intramembrane space and the matrix interact at the oxpho level. They can act as direct protonophores, shuttling hydrogen ions into the matrix (2,4-dinitrophenol is the classic example), or as ionophores, exchanging hydrogen ions for other mono- or divalent cations, or may generally increase the permeability of the inner membrane. The dissipation of the proton gradient without ATP generation can result in the generation of heat, and, in extreme conditions, a malignant hyperthermia syndrome can occur (32, 33). In this regard we can include most of the nonsteroidal anti-inflammatory drugs (NSAIDs) such as aspirin, diconfenac, and nimesulide [importantly, the pathogenesis of NSAID enteropathy also involves the uncoupling of mitochondrial oxphos, which alters the intercellular junction and increases intestinal permeability with consequent intestinal damage (63)], but also some antitumor drugs and antipsychotic, hypolipidemic, and antimitotic compounds. Interestingly, for all these drugs, the main structure-activity relationship is characteristically based on a lipophilic weak acid group (40, 77, 112).

Other xenobiotics may interfere with oxphos by direct inhibition of ATP synthase. The majority of these molecules are mycotoxins (such as oligomycin), but there are also well-known drugs such as propanolol, local anesthetics, and diethylstilbestrol. Intriguingly, some pharmacological activities of propanolol should be carefully reevaluated, especially with regard to their typical side effects such as cardiac output reduction.

Mitochondrial metabolic processes. Xenobiotics can interfere with catabolic and anabolic pathways in mitochondria. We described above some drugs that can induce dysfunction of the tricarboxylic acid cycle at the SDH and FH sites. In light of the etiopathogenic role of the congenital enzymopathic counterparts in serious forms of neurodegenerative diseases and cancer, the potential toxicological value of this dysfunction should not be underestimated.

In this respect, recent evidence reported by Nulton-Persson et al. (80) has shown that treatment with salicylic acid, and to a lesser extent, acetylsalicylate, increases the rate of uncoupled respiration in isolated cardiac mitochondria, in agreement with previous data (77). However, under the experimental conditions employed, loss of state 3 respiration resulted from inhibition of the tricarboxylic acid cycle enzyme α-ketoglutarate dehydrogenase. In particular, a kinetic analysis indicated that salicylic acid acts as a competitive inhibitor at the level of the α-ketoglutarate binding site. In contrast, acetylsalicylate inhibited the enzyme in a noncompetitive fashion, consistent with its interaction with the α-ketoglutarate binding site followed by enzyme-catalyzed acetylation. Furthermore, it was recently observed that cyclosporin reduced the concentrations of tricarboxylic acid cycle intermediates and inhibited mitochondrial oxphos at the level of ATP synthase in a time-dependent fashion (21). The real mechanism of such metabolic dysfunction is still being debated (i.e., energetic failure, reduced protein synthesis, or real enzymatic inhibition) (21). At first presentation, isoniazid overdosage can easily be mistaken for a case of diabetic ketoacidosis. Although the exact mechanism is still controversial, the inhibition of pyruvate conversion to lactate and the interference with NADH synthesis in the tricarboxylic acid cycle have both been suggested to contribute to the lactic acidosis observed in isoniazid toxicity. Intriguingly, serum isoniazid levels have not been helpful in the evaluation of isoniazid toxicity and treatment (1).

Another fundamental metabolic pathway that is often affected by xenobiotics is β-oxidation. Many drugs (tetracycline derivatives, NSAIDs such as ibuprofen and iroprofen, glucocorticoids, antidepressants such as amineptine and tianeptine, some statins, fibrates, estrogens, and some antiarrhythmics and antiangiinal drugs such as amiodarone and perhexiline) can directly and/or indirectly interfere with mitochondrial fatty acid oxidation, with important safety concerns, particularly with respect to the liver. However, the precise molecular mechanisms underlying this dysfunction have not been clearly established. The pathogenesis often appears secondary to an MRC derangement that heavily hampers NADH and/or FADH₂ oxidation (26, 32, 42). With respect to fibrates and thiazolidinediones, it is interesting to note that our data (90, 91), confirmed by Brunmair et al. (11, 12), showed that the dysfunction in glucose metabolism and/or β-oxidation significantly correlates with the level of complex I inhibition. Interestingly, Vickers et al. (110) recently showed a direct inhibition by etomoxir of the mitochondrial β-oxidation rate-limiting enzyme carnitine palmitoyltransferase I, which is associated with oxidative stress, inflammation, and apoptosis in the liver.

Mitochondrial protein synthesis. The close similarity between bacterial and mitochondrial ribosomes makes the latter a potential target for bacteriostatic antibiotics such as chloramphenicol, aminoglycosides, tetracycline, and the newest family, the oxazolidinones (73). For the latter class, a direct correlation has been demonstrated in both clinical and toxicological studies between the bacterial MIC₉₀, the IC₅₀ for mitochondrial protein synthesis, and the potential for mammalian toxicity, suggesting that this toxicity is manifested as a consequence of their effects on mitochondria (73, 112). In the four years since FDA approval of the oxazolidinone antibiotic linezolid, a number of papers have surfaced reporting lactic acidosis, peripheral and optic neuropathy, thrombocytopenia, and pure red cell aplasia resulting from prolonged use, which are all syndromes commonly associated with mitochondrial injury (73). It is worth noting that antibiotic inhibition of mammalian mitochondrial protein synthesis is often disregarded by not considering synergistic pharmacological interactions with other mitochondrial toxins.

Mitochondrial channels and mitochondrial permeability transition pores. Many well-known drugs (e.g., potassium channel openers such as nicorandil and diazoxide as well as antidiabetic and antitumor sulfonlyures) modify the activity of different mitochondrial channels, which have a fundamental role in maintaining the electrolyte homeostasis of mitochondria. Although these drugs are known to interact with components of various ion channels, the potential toxic effects and/or therapeutic applications of these mitochondrial channel alterations have not yet been clearly defined. Similar considerations should be made for drugs that interact with or modulate
components of the mitochondrial permeability transition pore (e.g., cyclosporine A binding of cyclophilin D, lonidamine binding of adenine nucleotide translocase, and drugs binding to the mitochondrial benzodiazepine receptor for which the putative toxic effects remain controversial) (6, 13, 23, 81, 37, 49).

The permeability transition pore is a high-conductance, nonspecific pore in the inner mitochondrial membrane composed of proteins that link the inner and outer mitochondrial membranes. When the permeability transition pore is opened as a result of exposure to high calcium or inorganic phosphate, depletion of NAD(P)H, alkaline pH, or ROS, low-molecular-weight substrates can freely penetrate the mitochondrial matrix, carrying along with them water and resulting in mitochondrial swelling and the release of cytochrome c into the cytosol. Cytochrome c release triggers a cascade of events that will lead to either apoptosis (in ATP-replete cells) or necrosis (in ATP-depleted cells). The toxicity of t-butyl-hydroperoxide and valproic acid and the chronic hepatotoxicity of diclofenac and other NSAIDs are mediated by this mechanism (8, 86). In general, the opening of this particular mitochondrial pore represents the common final event produced by numerous cell and mitochondrial toxins.

There are ongoing debates about drug actions and mitochondrial channels. For example, Skalska et al. (99) have proposed that sulfonlureas induce mitochondrial swelling, the lowering of Δψ, and an efflux of calcium from the matrix, mainly by activating the mitochondrial permeability transition. On the other hand, Fernandes et al. (36) hold that sulfonlureas interfere with mitochondrial bioenergetics mainly by permeabilizing the inner mitochondrial membrane to chloride ions and promoting a net chloride-potassium cotransport inside mitochondria.

Mitochondria and Drugs: Therapeutic Potential

Antioxidants. The first class of drugs developed specifically for the treatment of mitochondrial dysfunction are antioxidants, exemplified by 2,3-dimethoxy-5-methyl-6-decaprenyl-1,4-benzoquinone [coenzyme Q10 (CoQ10)], a fat-soluble quinone with a side chain of 10 isoprenoid units. Apart from its physiological electron carrier function, CoQ10 also seems to stabilize the MRC complexes and acts as a potent scavenger of oxygen free radicals. Accordingly, CoQ10 has been applied therapeutically in different congenital oxphos diseases (19, 64, 88, 120). Interestingly, some positive effects have been also reported in neurodegenerative disorders in general and in Alzheimer disease in particular, although these results have not been confirmed (19, 64, 120). In addition, menadione and phylloquinone (vitamin K compounds, which are well-known uncouplers of the electron respiratory chain) alone or in conjunction with ascorbate have been adopted to treat various mitochondrial disorders (e.g., cyclophilin A binding of cyclophilin D, lonidamine binding of adenine nucleotide translocase, and drugs binding to the mitochondrial benzodiazepine receptor for which the putative toxic effects remain controversial) (6, 13, 23, 81, 37, 49).

The electron respiratory chain may be affected by three mechanisms: 1) directly by alteration of a single complex [rotenone and analogs and arsenic trioxide (which partially inhibits) for complex I (2, 94); tamoxifen for complex III and IV (75); and genistein and 17α- or β-estradiol for ATP synthase (3, 71)]; 2) indirectly by generation of free radical species by the same agents that disrupt the electron respiratory chain, by impairing complex I and III (82); and 3) indirectly via photosensitizers or inhibitors of the intrinsic antioxidant defenses of mitochondria (i.e., the superoxide dismutase inhibitor 2-methoxyestradiol) (29).

Mitochondrial permeability transition pores may be affected by interfering with the pores’ physiological functions [i.e.,

Unfortunately, the molecular structure of mitoKATP channels is not well known, in contrast to sKATP channels, which are composed of a pore-forming subunit (Kir6.1 or Kir6.2) and a sulfonylurea receptor (SUR1, SUR2A, or SUR2B). Recently, it has been observed that some drugs behave as potassium channel openers capable of acting at the level of cellular membranes, including mitochondrial membranes (80). The pharmacological activity of such drugs can therefore also be ascribed to mitochondrial ion modulation. These compounds, which include cromakalim (47), nicorandil (23, 57), and pinacidil (28, 67), were found to modulate K1 channels both in smooth muscle cell membranes and in mitochondria displaying antianginal (nicorandil) or antihypertensive (cromakalim and pinacidil) profiles. Therapeutic activity at the mitochondrial level has also been reported for a variety of older antihypertensive agents, notably diazoxide and minoxidil sulfate, which may also influence the activity of mitoKATP channels (13, 31). Moreover, diazoxide was recently shown to decrease succinate oxidation in a dose-dependent manner (31, 48, 114), albeit at higher concentrations than those necessary to activate mitoKATP. Thus it has been proposed that the cardioprotective effects of diazoxide may result from the inhibition of SDH and a decrease in respiration, rather than from the opening of mitoKATP channels (48). Consistent with the findings that SDH is part of a protein complex capable of transporting K+, SDH may also regulate mitoKATP by means of its physical interaction with the ionophore rather than via its role in oxphos (81).

Such data, once confirmed, could offer new potential therapeutic strategies for different degenerative diseases and stress the potential therapeutic role of mitoKATP modulation.

Anticancer agents. Another fundamental pharmacological area in which the interaction between drugs and mitochondria could have striking possibilities is cancer therapy. On one hand, the physicochemical and biological properties of mitochondria expose them to toxic agents, but on the other hand, the same properties could allow us to consider mitochondria as a target of chemotherapeutic agents for selective anticancer therapy (27, 30, 45, 89, 116).

Mitochondrial dysfunction could trigger pathways capable of inducing cell apoptosis or necrosis. Several molecular mechanisms involved in mitochondrial toxicity by different xenobiotics can be and/or have already been utilized in cancer therapy. The following mechanisms are promising mitochondrial therapeutic targets.

mtDNA biogenesis may be affected by inhibiting topoisomerase II (etoposide and analogs, but also cisplatin and 5-fluorouracil and their analogs) or poly-γ (as noted above in the case of antiviral nucleoside analogs) (16, 20, 46, 86, 103).

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AJP-Cell Physiol • VOL 293 • JULY 2007 • www.ajpcell.org

Invited Review

MITOCHONDRIAL PHARMACOTOXICOLOGY

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Potassium channel opening can be affected by analogs of dequalinium, diazoxide, and amiodarone, which increase the permeability of the mitochondrial membrane to protons or potassium and induce a decrease in $\Delta \psi$, mitochondrial swelling, decrease in ATP synthesis, and release of cytochrome c (54, 87, 108).

Inhibition of Bcl-2/Bcl-X$_L$ or activation of Bax/Bak by antisense Bcl-2/Bcl-X$_L$ or by a single-chain antibody can sensitize apoptosis-resistant cancer cells to chemotherapy (5, 62, 118).

Importantly, the latter two classes of anticancer mechanisms represent interesting and innovative therapeutic approaches in which mitochondria are the primary targets. However, these are still in preclinical testing phases, and the speculated applications in cancer and the real therapeutic index must be accurately evaluated. Moreover, independent of the strategy adopted to induce apoptosis and/or necrosis in cancer cells, the best possible targeting of drugs, first to neoplastic cells and then to their mitochondria, must be ensured. As already stated, some authors (8, 79, 116) have suggested that positively charged amphiphatic molecules can be attracted by and penetrate into mitochondria in response to the highly negative membrane potential and even more so in neoplastic cells, which present a more elevated plasma/mitochondrial membrane potential than differentiated cells. Currently, this strategy represents the best compromise to target proapoptotic drugs specifically to cancer cells. However, it should be kept in mind that the in vivo biological environment is extremely variable in cancer; thus patients may be exposed to dangerous and dramatic side effects.

Mitochondria and Drugs: Perspectives

The future of mitochondrial pharmacology appears to be headed first toward the development of therapies for glucidic and lipidic metabolism and energy expenditure disorders. Indeed, recent experimental and clinical research has focused on molecular dysfunction at the mitochondrial level for the pathogenesis of some inherited and acquired metabolic diseases (i.e., some mitochondrial forms of non-insulin-dependent diabetes mellitus, metabolic syndrome, hyperlipoproteinemia). These data have been confirmed by experimental evidence showing that it is possible to modulate glucose and/or fatty acid oxidation pharmacologically at the cellular level (32, 79, 102).

Modulating the expression and/or activity of the so-called uncoupling proteins in different tissues in general, or of adipose tissue in particular, could represent a new and revolutionary approach to the pharmacological treatment of obesity (51). Interestingly, new and more potent peroxisome proliferator-activated receptor (PPAR) ligands (especially type $\delta$) are being developed for this specific clinical indication that exploit the capability to induce the expression of genes required for fatty acid catabolism and adaptive thermogenesis (51, 115). Given the adverse side effects of some PPAR ligands, an accurate analysis of the potential interactions with mitochondria is imperative (68).

Moreover, recent data indicate that mitochondria in cancer not only represent mere effectors of apoptosis but also have a more complex role in oncogenesis and oncosuppression (30, 44, 86, 90, 91). Additional findings (91, 104) have indicated that electron respiratory chain dysfunction can induce differentiation in different human neoplastic cell lines, suggesting that mitochondria may play additional roles in regulating cell homeostasis. Finally, modulation of the activity of the mitochondrial electron respiratory chain by so-called NO-releasing drugs may expand the potential therapeutic applications in mitochondrial medicine. As these therapies are developed and tested, it will be important for us to beware of causing undesired toxic effects derived from drug-mitochondria interactions that may not have been carefully considered (76, 89).

REFERENCES


