Software for simulating calcium-triggered exocytotic processes

Germán Carrera,1 Amparo Gil,1 Javier Segura,1 and Bernat Soria2

1Departamento de Matemáticas, Estadística y Computación, Universidad de Cantabria, Santander; and
2CABINER, Américo Vespucio S/N, Seville, Spain

Submitted 20 February 2006; accepted in final form 26 August 2006

Carrera G, Gil A, Segura J, Soria B. Software for simulating calcium-triggered exocytotic processes. Am J Physiol Cell Physiol 292: C749–C755, 2007. First published August 30, 2006; doi:10.1152/ajpcell.00082.2006.—We describe a software package for the simulation of exocytotic events from readily releasable pools of secretory vesicles in neuroendocrine cells and presynaptic terminals. The visual package Ca3D_Exolab simulates the entry of Ca2+ through the calcium channels, the kinetic reactions of calcium with buffers, the diffusion of calcium and mobile buffers, and the kinetic reactions of calcium with the secretory vesicles. The location of both channels and secretory vesicles can be set by using a graphical interface. Calcium and buffer concentrations at different depths from the cellular membrane and capacitance time courses are obtained as outputs. The software package also provides a descriptive statistical data analysis of the different output data.

THE QUANTITATIVE MODELING of calcium dynamics and related processes is a useful tool for confronting theory with experimental studies of the secretory machinery in neuroendocrine cells and presynaptic terminals. Most computational models of calcium dynamics in cells are based on the solution of differential equations assuming, for instance, a spherical model of the cell with uniform time-dependent distributions of calcium below the membrane (see, for instance, Refs. 7–9 for related software). Although these models provide an average description of the calcium processes, it is extremely difficult to build reliable simulations based on differential equations for describing aspects related to the geometry of the system. For instance, the influence of the spatial distribution of the calcium channels on the calcium and buffer time courses is difficult to model with a differential approach; a spatial organization of the calcium channels in clusters and a random distribution of channels will lead, in general, to very different calcium time courses.

When the secretory machinery of vesicles enters into play, the influence of the geometrical aspects becomes even more important; in this case, the relative position of the calcium channels (organized or not in clusters) and the secretory sensors of the vesicles are crucial parameters of the system. To incorporate information on the geometry of the system, the use of microscopic models, such as those based on Monte Carlo (MC) methods (see Ref. 2, for example), is particularly appropriate. However, because of the complexity of the algorithms involved, MC methods are probably not as popular as differential methods. A “raw” MC code is not an easy thing to use, whereas many mathematical packages are equipped with friendly differential equation solvers. In trying to overcome this limitation, we have built a user-friendly software environ-

THE MATHEMATICAL MODELS

The mathematical models behind Ca3D_Exolab were introduced, analyzed, and explained in detail in previous publications (2, 10). We now summarize the main ingredients of the model.

Domain of Simulation

A three-dimensional (3-D) orthogonal grid maps the (by now) two possible choices of the domain for the simulations: a conical domain and a cylindrical domain. The size $\Delta x$ of the lateral sides of the cubic compartments of the grid can be selected by the user of the program. This size represents the spatial resolution for the simulations.

The choice of a conical domain is appropriated for simulating secretory events in spherical cells, assuming that the portion of the membrane represented by the base of the cone is embedded inside a larger portion of the membrane where the calcium channels are, on average, uniformly distributed. In this way, one can consider that the net flux of ions and buffers through the lateral sides of the cone is zero. This approximation is also safe for smoothly nonuniform configurations of channels in which the conical domain is placed inside a region where the distribution of channels is sufficiently uniform. The choice of a cylindrical domain seems appropriated for simulating calcium processes in cilliar cells or presynaptic terminals.

Calcium Current

We consider a simple model for calcium influx in which the calcium current is constant during a pulse. The selection of
different shapes of the current and the stochastic modeling of channel gating will be considered in future versions of the software.

**Diffusion of Mobile Particles**

For modeling the 3-D diffusion of calcium and mobile buffers, we use a random walk algorithm (2). In this way, calcium ions and mobile buffers move in intervals of time:

\[ \Delta t = \frac{\Delta x^2}{4D}, \]

where \( \Delta x \) is the spatial resolution for the simulations and \( D \) is the diffusion coefficient of the mobile particle.

**Secretory Vesicles and Buffers**

For simulating the kinetic reactions of the secretory sensors of vesicles and buffers with calcium, we consider a discrete (stochastic) modeling (2, 10). For the secretory vesicles, a noncooperative scheme with a variable number of binding sites \( n \) is considered:

\[
\begin{align*}
X &\xrightarrow{k_{on}[Ca]} nXCa_1 \\
&\xrightarrow{2k_{off}} \cdot \cdot \cdot \\
&\xrightarrow{p n k_{off}} XCa_n \rightarrow F
\end{align*}
\]

where \( X \) represents the calcium sensors of the secretory vesicles, \( k_{on} \) is the on rate of the reaction, \( k_{off} \) is the off rate of the reaction, and \( p \) is the fusion rate. The kinetic reactions of calcium ions to binding sites (of both buffers and secretory vesicles) are described by first-order equations:

### Table 1. Parameters used in the case study of secretion in chromaffin cells

<table>
<thead>
<tr>
<th>Geometrical parameters</th>
<th>Vesicle sensor</th>
</tr>
</thead>
<tbody>
<tr>
<td>Choice of conical domain</td>
<td>Calcium channel</td>
</tr>
<tr>
<td>Radius</td>
<td>Calcium channel</td>
</tr>
<tr>
<td>Depth</td>
<td>Calcium channel</td>
</tr>
<tr>
<td>Length of the grid</td>
<td>Calcium channel</td>
</tr>
<tr>
<td>Distribution of channels specified by the user</td>
<td>Calcium channel</td>
</tr>
<tr>
<td>Number of calcium channels</td>
<td>Calcium channel</td>
</tr>
<tr>
<td>Distribution of vesicles specified by the user</td>
<td>Calcium channel</td>
</tr>
<tr>
<td>Number of vesicles</td>
<td>Calcium channel</td>
</tr>
</tbody>
</table>

**Calcium current parameters**

- **Basal concentration** \( [Ca^{2+}]_0 \) = 0.1 \( \mu \)M
- **Diffusion coefficient** \( D_{Ca} \) = 220 \( \mu \)m²/s

**Vesicles**

- **Number of binding sites** \( n_{bs} \) = 5
- **Forward binding rate** \( k_1 \) = \( 3 \times 10^{8} \) M⁻¹s⁻¹
- **Dissociation constant** \( K_d \) = 10 \( \mu \)M
- **Fusion rate** \( FR \) = 40,000 s⁻¹

**Endogenous buffer**

- **Total concentration** \( [EFB] \) = 500 \( \mu \)M
- **Forward binding rate** \( k_1 \) = \( 5 \times 10^{9} \) M⁻¹s⁻¹
- **Dissociation constant** \( K_d \) = 10 \( \mu \)M

**Exogenous buffer fura-2**

- **Total concentration** \( [Fura-2] \) = 100 \( \mu \)M
- **Forward binding rate** \( k_1 \) = \( 5 \times 10^{8} \) M⁻¹s⁻¹
- **Dissociation constant** \( K_d \) = 0.24 \( \mu \)M
- **Diffusion coefficient** \( D_{Fura-2} \) = 42 \( \mu \)m²/s

**Calcium current parameters**

- **Basal concentration** \( [Ca^{2+}]_0 \) = 0.1 \( \mu \)M
- **Diffusion coefficient** \( D_{Ca} \) = 220 \( \mu \)m²/s

**Vesicles**

- **Number of binding sites** \( n_{bs} \) = 5
- **Forward binding rate** \( k_1 \) = \( 3 \times 10^{8} \) M⁻¹s⁻¹
- **Dissociation constant** \( K_d \) = 10 \( \mu \)M
- **Fusion rate** \( FR \) = 40,000 s⁻¹

**Endogenous buffer**

- **Total concentration** \( [EFB] \) = 500 \( \mu \)M
- **Forward binding rate** \( k_1 \) = \( 5 \times 10^{9} \) M⁻¹s⁻¹
- **Dissociation constant** \( K_d \) = 10 \( \mu \)M

**Exogenous buffer fura-2**

- **Total concentration** \( [Fura-2] \) = 100 \( \mu \)M
- **Forward binding rate** \( k_1 \) = \( 5 \times 10^{8} \) M⁻¹s⁻¹
- **Dissociation constant** \( K_d \) = 0.24 \( \mu \)M
- **Diffusion coefficient** \( D_{Fura-2} \) = 42 \( \mu \)m²/s

**Fig. 1.** Schematic representation of the location of vesicles below the membrane. In this representation, a configuration of 5 binding sites per vesicle is considered. Calcium channels also are shown.

**Fig. 2.** A: screen shot for the graphical interface of Ca3D_Exolab for setting the position of the channels. B: screen shot for setting the position of the vesicles. The configurations in A and B correspond to the simulated case study of chromaffin cells.
\[
d\frac{[\text{Ca}]}{dt} = \sum_i k_{i}^{\text{off}}[\text{CaB}_i] - k_{i}^{\text{on}}[\text{Ca}][\text{B}_i],
\]
\[
\sum_i d[B_i]/dt = d[\text{Ca}]/dt, d[\text{CaB}_i]/dt = -d[B_i]/dt
\]

where \([\text{Ca}],[\text{B}_i] \text{ and } [\text{CaB}_i]\) are, respectively, the concentrations of calcium, unbound buffer, and bound calcium. In our simulation scheme, these equations are interpreted stochastically, evaluating the probabilities associated with each of the processes and generating events according to such probabilities (2).

The spatial distribution of the binding sites of buffers is randomly and uniformly distributed over the 3-D domain, whereas the binding sites of the secretory vesicles are spatially restricted to the location of the vesicles. The mechanism for secretion is simulated by placing the number of binding sites, initially free, corresponding to the kinetic scheme considered, in each of the submembrane compartments where the protein lies. In Fig. 1, a schematic representation of the location of vesicles below the membrane is shown. In this representation, a five-binding sites kinetics scheme per vesicle is considered. The calcium channels are also shown.

In our model, the fate of each individual vesicle is recorded. The finite size of the vesicles is neglected: we simulate the kinetics for vesicle fusion, but we do not consider the fact that calcium ions and buffer molecules are not free to diffuse through the space occupied by the vesicles of the readily releasable pool (RRP). This is justified given the small number of vesicles that typically constitute the RRP.

Replenishment of the RRP is not considered in this model. The simulations are restricted to trains of pulses in the subsecond range. Endocytosis and the mobilization of additional vesicles can then be thought to be too slow to be noticed.

---

Fig. 3. Screen shots of the output data generated by Ca3D_Exolab. A: average \([\text{Ca}^{2+}]\) time course (0 to 50 nm from the cellular membrane) for the case study of chromaffin cells. B: time course of the number of accumulated secretory events.

Fig. 4. Screen shots of the output data generated by Ca3D_Exolab. The plot corresponds to the average free buffer concentrations \([\text{EFB}]_{\text{free}}\) (top left) and \([\text{Fura-2}]_{\text{free}}\) (top right) (0 to 50 nm from the cellular membrane) for the case study of chromaffin cells.
DESCRIPTION OF THE SOFTWARE

Using the theoretical ingredients described above, we have built a software package (Ca3D_Exolab) for simulating secretory events in neuroendocrine cells and presynaptic terminals. The package can be downloaded at http://personales.unican.es/gila/bbva.html.

The MC code for simulating calcium dynamics that, in an improved version, form the core of Ca3D_Exolab, was extensively tested, and its results were proved to match with theoretical expectations at long times (2). Explicit tests for the routines implementing the random walk algorithm and the kinetic reactions of calcium with buffers can be found in Ref. 3.

The outputs of Ca3D_Exolab are calcium and buffer concentration time courses at different depths from the cellular membrane and the number of secretory events as a function of time. For the different output data, a descriptive statistical data analysis is available.

Two Illustrative Examples

As a practical example, consider two different case studies of secretion: chromaffin cells (see Ref. 11, for example) and calyx of Held (see Refs. 5, 6).

For chromaffin cells, differential methods are generally able to describe the time course of average calcium concentrations below the membrane. Concentrations are high enough so that the averages make sense, and so does the differential modeling of these average values. When high concentrations occur, the MC method implemented uses average descriptions when suitable (see Refs. 2 and 3 for further details).

The calyx of Held is a good example showing when it is not so clear that average descriptions are valid, as commented in Ref. 6. Indeed, calcium concentrations are so low that, for the spatial resolutions considered, very few or no calcium ions may be found, as the results show. This fact, together with the fact that the binding of calcium to vesicle sensors also affects calcium concentration, poses a serious limitation on differential models, which are overcome by the MC approach.

Chromaffin Cells

We followed descriptions in Refs. 2 and 10 for modeling of these cells. An appropriate domain for simulating localized submembrane time courses is a conical domain. We chose, for example, a cone with a 1-μm radius of base (representing the radius of a portion of the membrane) and 5 μm in height. The spatial resolution Δx was set to 50 nm.

Initial conditions and reaction-diffusion parameters. We considered the following input data for the initial conditions and reaction-diffusion parameters (2, 10): for calcium, initial concentration ([Ca$$^{2+}$$]_{initial}) = 0.1 μM, and diffusion coefficient ($$D_{Ca^{2+}}$$) = 220 μm$$^2$/s; for the endogenous fixed buffer (EFB), total concentration ([EFB]_{total}) = 500 μM, affinity ($$K_{d}$$(EFB)) = 10 μM, and forward Ca$$^{2+}$$ binding rate ($$k_{+}$$) = 5 × 10$$^{8}$$ M$$^{-1}$$s$$^{-1}$; for the exogenous mobile buffer fura-2, total concentration...
Calcium current parameters

\[ \text{m}^2/\text{s}, \quad \text{H}^9262 ([\text{fura-2}]_{\text{total}}) \]

Geometrical parameters


Calvin current parameters

Number of calcium pulses
Frequency at which the pulses are delivered
Duration of the simulation
Unitary current per channel

Kinetic parameters

Calcium
Basal concentration
Diffusion coefficient

Vesicles
Number of binding sites
Forward binding rate
Dissociation constant
Fusion rate

Endogenous buffer
Total concentration
Forward binding rate
Dissociation constant

Exogenous buffer ATP
Total concentration
Forward binding rate
Dissociation constant

Diffusion coefficient

Table 2. Parameters used in the case study of secretion in the calyx of Held

<table>
<thead>
<tr>
<th>Geometrical parameters</th>
<th>Calcium current parameters</th>
<th>Kinetic parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td>Choice of cylindrical domain</td>
<td>Number of calcium pulses</td>
<td>Calcium</td>
</tr>
<tr>
<td>Radius</td>
<td>Frequency at which the pulses are delivered</td>
<td>Basal concentration</td>
</tr>
<tr>
<td>Depth</td>
<td>Duration of each pulse</td>
<td>Diffusion coefficient</td>
</tr>
<tr>
<td>Length of the grid</td>
<td>Diffusion coefficient</td>
<td>Vesicles</td>
</tr>
<tr>
<td>Choice of distribution of channels specified by the user</td>
<td>Duration of the simulation</td>
<td>Number of binding sites</td>
</tr>
<tr>
<td>Number of calcium channels</td>
<td>Unitary current per channel</td>
<td>Forward binding rate</td>
</tr>
<tr>
<td>Choice of distribution of vesicles specified by the user</td>
<td></td>
<td>Dissociation constant</td>
</tr>
<tr>
<td>Number of vesicles</td>
<td></td>
<td>Fusion rate</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Vesicles</th>
<th>Endogenous buffer</th>
<th>Exogenous buffer ATP</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of binding sites</td>
<td>Total concentration</td>
<td>Total concentration</td>
</tr>
<tr>
<td>Forward binding rate</td>
<td>Forward binding rate</td>
<td>Forward binding rate</td>
</tr>
<tr>
<td>Dissociation constant</td>
<td>Dissociation constant</td>
<td>Dissociation constant</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Initial conditions and reaction-diffusion parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td>([\text{Ca}^{2+}]_0 = 0.05 \mu\text{M})</td>
</tr>
<tr>
<td>(D_{\text{Ca}} = 220 \mu\text{m}^2/\text{s})</td>
</tr>
</tbody>
</table>

([fura-2]_{total}) = 100 \mu\text{M}, diffusion coefficient \((D_{\text{Fura-2}}) = 42 \mu\text{m}^2/\text{s}, K_d = 0.24 \mu\text{M}, \text{and } k_+ = 5 \times 10^8 \text{M}^{-1} \cdot \text{s}^{-1}\); and for the kinetic release model for vesicles, there were five independent binding sites, where \(K_d = 10 \mu\text{M}\) and \(k_+ = 3 \times 10^8 \text{M}^{-1} \cdot \text{s}^{-1}\). In addition, we considered a fusion rate of \(p = 40,000 \text{s}^{-1}\). The parameters of the system are summarized in Table 1.

**Topography of channels and vesicles.** Let us, for example, consider two clusters of channels (Fig. 2A) and a RRP composed of three vesicles (Fig. 2B). For this topographical configuration of channels and vesicles, typical outputs of Ca3D_Exolab are shown in Figs. 3 and 4. Figure 3A shows the average \([\text{Ca}^{2+}]\) time course from 0 to 50 nm from the cellular membrane.

Figure 3B shows the time course of the number of accumulated secretory events. In Fig. 4, the time courses of the average free endogenous and exogenous buffer (fura-2) concentrations (from 0 to 50 nm from the cellular membrane) are shown.

**Calyx of Held**

The parameters for the simulation of the calyx of Held presynaptic terminal were taken from Ref. 5. Because the release topography of the system cannot be inferred from experimental studies, the modeling can be used for testing different topographies.

**Domain of simulation.** In the case of the calyx of Held, we chose as domain for the simulations a cylindrical domain of 400 nm in height and 130 nm in radius (5). This last parameter corresponds approximately to the mean radius of the active zones in the presynaptic membrane, which are the regions of interest.

**Initial conditions and reaction-diffusion parameters.** We considered the following input data for the initial conditions and reaction-diffusion parameters (5): for calcium, \([\text{Ca}^{2+}]_{\text{rest}} = 50 \text{nM}\) and \(D_{\text{Ca}} = 220 \mu\text{m}^2/\text{s}\); for endogenous fixed buffer, \([\text{EFB}]_{\text{total}} = 80 \mu\text{M}\), \(K_d = 2 \mu\text{M}\), and \(k_+ = 5 \times 10^8 \text{M}^{-1} \cdot \text{s}^{-1}\); for ATP, total concentration \([\text{ATP}]_{\text{total}} = 580 \mu\text{M}\), diffusion coefficient \((D_{\text{ATP}}) = 220 \mu\text{m}^2/\text{s}, K_d = 200 \mu\text{M}, \text{and } k_+ = 5 \times 10^8 \text{M}^{-1} \cdot \text{s}^{-1}\); and for the kinetic release model for vesicles, there were five independent binding sites, where \(K_d = 10 \mu\text{M}\) and \(k_+ = 3 \times 10^8 \text{M}^{-1} \cdot \text{s}^{-1}\). In addition, we considered a fusion rate of \(p = 40,000 \text{s}^{-1}\).

**Topography of channels and vesicles.** As mentioned previously, the release site topography at most synapses, including the calyx of Held, cannot be measured experimentally. According to the theoretical study in Ref. 5, at each active zone, the Ca\(^{2+}\) that controls phasic release is supplied by 10 or more calcium channels. These channels are grouped into one or a few clusters, with a channel cluster covering an area of <50 nm across. Let us consider a RRP composed of three vesicles associated to this active zone.

![Fig. 7. Screen shots of the output data generated by Ca3D_Exolab. The plot corresponds to the time course of the number of accumulated secretory events for the case study of the calyx of Held with the vesicle configuration shown in Fig. 5A.](http://ajpcell.physiology.org/ by 10.220.33.1 on November 28, 2017)
Fig. 8. Screen shots of the output data generated by Ca3D_Exolab. The plot corresponds to the average [Ca\textsuperscript{2+}] time course (0 to 10 nm from the cellular membrane) for the case study of the calyx of Held. The descriptive statistical summary of the data is also shown.

Fig. 9. Screen shots of the output data generated by Ca3D_Exolab. The plot corresponds to the average free buffer concentrations [EFB]\textsubscript{free} (top left) and [ATP]\textsubscript{free} (top right) (0 to 10 nm from the cellular membrane) for the case study of the calyx of Held.
We simulated the secretory response of the RRP under two different topographical configurations of vesicles (Fig. 5, A and B) and a topographical configuration of channels consisting in a single cluster of 12 channels (Fig. 6). The spatial resolution $\Delta r$ was set to 10 nm.

In Table 2 we summarize the set of input parameters used in the simulations. For simplicity, we assumed a constant current pulse per channel of 0.02 pA and a duration of 1 ms. For the topographical configuration of vesicles shown in Fig. 5A, typical outputs of Ca3D_Exolab are shown in Figs. 7–9. Figure 7 shows the time course of the number of accumulated secretory events; Fig. 8 shows the average $[\text{Ca}^{2+}]$ time course from 0 to 10 nm from the cellular membrane. The descriptive statistical data analysis for these data is also shown. In Fig. 9, the time courses of the average free endogenous buffer and ATP concentrations (from 0 to 10 nm from the cellular membrane) are shown.

A different secretory scenario appeared when the topographical configuration of vesicles in Fig. 5B was considered; in this case, no secretory events were recorded during the simulation time. This illustrates the necessity of being able to simulate different topographies for obtaining information on the feasibility of a given set of kinetic and diffusive parameters. Average descriptions will fail to provide this type of information.

GRANTS

We acknowledge financial support from Fundación BBVA under the project “Understanding the secretory machinery of living cells: mathematical models and computational tools.” A. Gil acknowledges financial support from the Ministerio de Educación y Ciencia (Programa Ramón y Cajal).

REFERENCES